

# Supplementary Information

## Expansion of the mutually exclusive spliced exome in *Drosophila*

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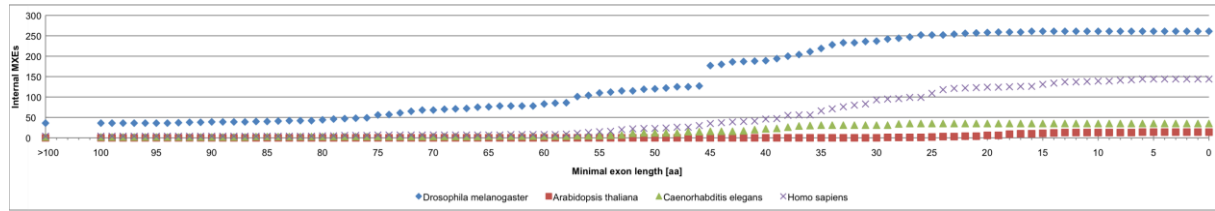
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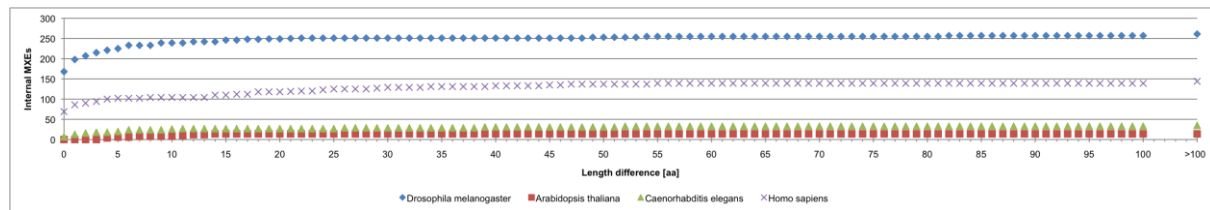
(email: mako@nmr.mpibpc.mpg.de)

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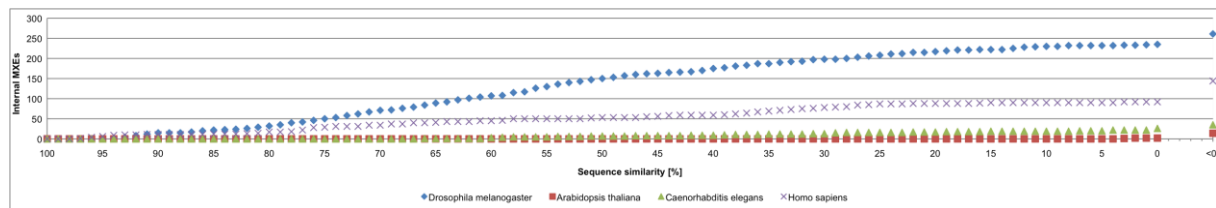
# 1 Supplementary Figures



**Supplementary Figure S1. Number of annotated internal mutually exclusive spliced exons (MXEs) as function of the respective length of the MXE.** The two noticeable jumps in the scatter plot of the *Dm* MXEs are due to the MXEs in the large clusters of the DSCAM gene. The shorter the exons are the more probable it becomes that their sequences are featureless and that false positive candidates will be predicted. Therefore, we introduced a parameter “minimal exon length”. Based on the analysis of all annotated MXEs we set this parameter to 15 residues.

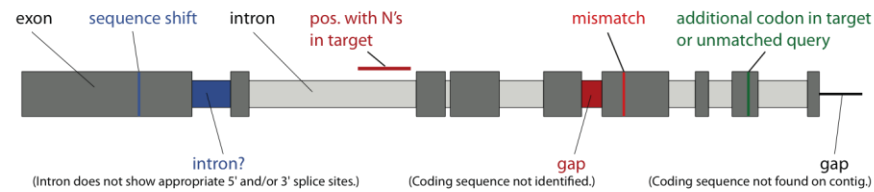


**Supplementary Figure S2. Number of annotated internal mutually exclusive spliced exons (MXEs) as function of the minimal length difference to another MXE of the same cluster.** To determine a suitable cut-off for the length difference in the search we analysed all internal clusters of annotated MXEs in the *Drosophila melanogaster* genome (*Dm*, Flybase release 5.36). To exclude that the determined characteristics are *Drosophila* specific we also analysed the annotated mutually exclusive exomes of *Homo sapiens* (*Hs*, NCBI release 37.3), *Caenorhabditis elegans* (*Ce*, WormBase release WS230), and *Arabidopsis thaliana* (*At*, TAIR release 167). These species have been chosen because of their widespread taxonomic distribution and their advanced and detailed annotations. For all species analysed the curves look very similar. 64%, 20%, 48% and 0% of the annotated MXEs of *Dm*, *Hs*, *Ce*, and *At*, respectively, have no length difference (86%, 71%, 57% and 43% have length difference of less than five residues). Therefore, a cut-off for the length difference of 20 residues should be appropriate to reconstruct almost all annotated cases and to not include too many mispredictions (95%, 82%, 77% and 100% have length difference of less than 20 residues).

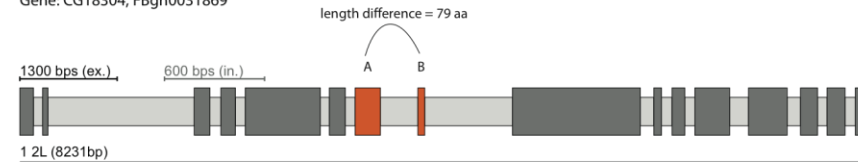


**Supplementary Figure S3. Number of annotated internal mutually exclusive spliced exons (MXEs) as function of the sequence similarity to another MXE of the same cluster.** In the case of similarity, two slightly different similarity scores can be calculated for a pair of MXEs dependent of which has been used as reference. Here, we included the respective higher scores. In this project, we were supposing that the MXEs of a cluster code for identical secondary structural elements of the protein like in the *Dm* muscle myosin heavy chain. If this condition holds true the MXEs should show a certain degree of sequence similarity. Analysis of the MXEs of *Dm* shows that 94.9% of the MXEs, which show any sequence similarity, have a sequence similarity of more than 15%. In *Hs* and *Ce*, 98% and 86% of the MXEs, which show any sequence similarity, have higher sequence similarities than 15%. Therefore, we decided to use 15% sequence similarity as cut-off for further predictions. However, a few cases of annotated MXEs do not show any sequence similarity and can not be reconstructed with our method (see difference of the two rightmost numbers).

## Legend



Gene: CG18304, FBgn0031869



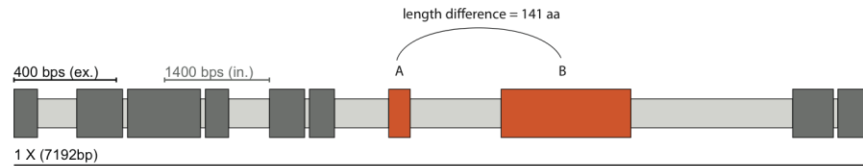
For clarity introns have been scaled up by a factor of 2.23

	10	20	30	40	50	60	70	80
exonA	LQRLRDREKRRVRFSCGTQTEVPLEVVAFFRGQTQVATVQSDMSTSVENLVTSNVAVTQTDTEFVDPDRNVSIERETMSSEFF							
exonB	---AEHLRKKVTRFEDENESLMMQLKKMATRSR							

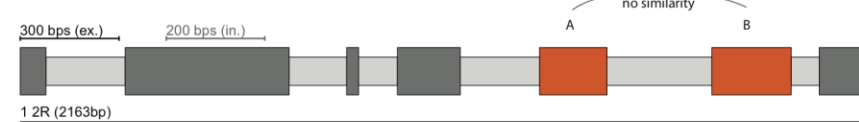
	90	100	110
exonA	AGLFPFSSSRVQSGSLLFSAISHVLLSGA		
exonB			

Gene: PhKgamma, Phosphorylase kinase  $\gamma$ , FBgn0011754



For clarity introns have been scaled down by a factor of 3.42

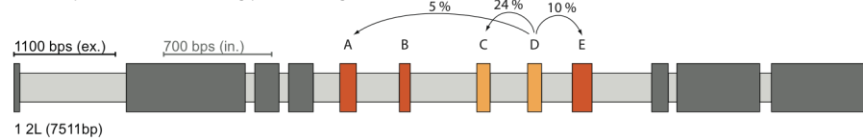
Gene: endoB, endophilin B, FBgn0034433



For clarity introns have been scaled up by a factor of 1.49

	10	20	30	40	50	60	70	80
exonA	MQHPKPLRLINSEDDCGPSSVSMHSCDSLSLGGIALDSDFDPLDKSLTNLLEDFHIEFDTTAVST							
exonB	LGGPTPIYIFLDVNEASAKSINSISGAARGPNNHSAANMAATGHKPNQPMHVSTQMQRARVLCSDAKDHTLNLSANE							

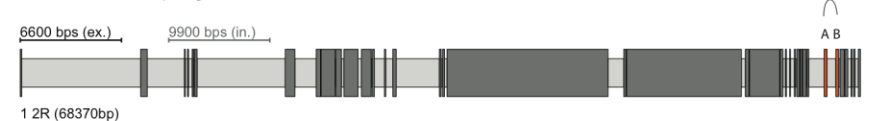
Gene: TepII, Thiolester containing protein II, FBgn0041182



For clarity introns have been scaled up by a factor of 1.67

	10	20	30	40	50	60	70	80
exonA	-----EFPDYVEDDFEIYAFENNLDALPMPAIANFPDPTGNTVQP-VEIRKNFADVWIWQSIGRS-							
exonB	-----AKAIPESLDYQV-----EDSIS-YDEVDAISITSSTKIEL-----VRTNFAEVWMWTTSDNGS							
exonC	-----EFPYIAF-----SLAAPAAIAGMPTSSIASHPNQA-PQ-----IRKEFFENWIFNYAEN--							
exonD	-----ERRIFIRP-----GIGFPRFLFNRVTVAGSLPPNVIPE-PQ-----VRKEFFENWIFNIFEN-							
exonE	GPLVMSYVFE-GSRHPWITRPRYRVGIRG-----DSGDRISFLSQSLNDRNLKEILLKQTPQRTIRKEFFETWFFEN-----							

Gene: shot, short stop, FBgn0013733



For clarity introns have been scaled down by a factor of 1.50

	10	20	30	40	50	60
exonA	AKGRTNIELREQFILADGVSQSMAAFTPRRSTPNAAATASSSPHAHNGGSSNLPFYMSSGQGPPIIK					
exonB	AD-EHLAELMFIKIRAQDQVFCAPPFIH-----MGAGGTVFVRCNTRSRSVFLSPHVLHCHPTTHW					

Gene: Dscam, Down syndrome cell adhesion molecule, FBgn0033159



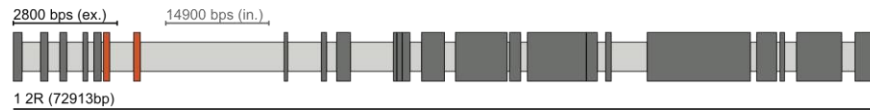
For clarity introns have been scaled down by a factor of 1.47

1600 1610 1620 1630 1640 1650 1660

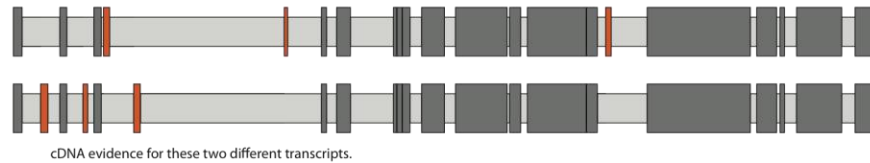
exon17A GTIAPSRDLPELSAEDTIRIILS-----NLNLVVFVVAALLVIIIAIIVICILRSKGN--HHK

exon17B GTIAPLDDGSGHGNVHTRIRLPAWMEWLDLNFVMPLIATVVVAVGICVVCVALSRRRADDMR

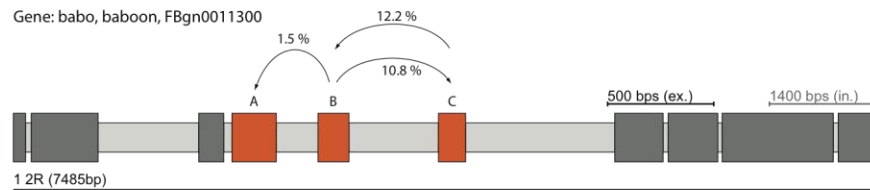
Gene: Nipped-A, FBgn0053554



For clarity introns have been scaled down by a factor of 5.36



Gene: babo, baboon, FBgn0011300



For clarity introns have been scaled down by a factor of 2.78

10 20 30 40 50 60 70

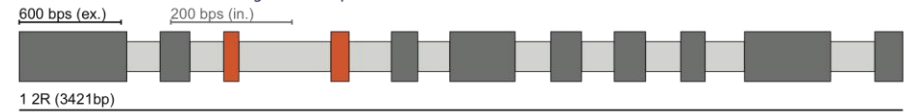
exonA CLHKSQIFPGRSIWCNDLGHGGPTARVGRNGAHACKDRDFCN-REFLWPKTKDQSRDVEEGRQISVQ

exonB CMVVKYNMQRSK-----PFECLTSNERFDYRIDCKS-DFCNKNEIMKRI FET

exonC CITDQLP-----PEDPTSCKLNSEAGSSQCCAE-DFCNTRNYSVGLP

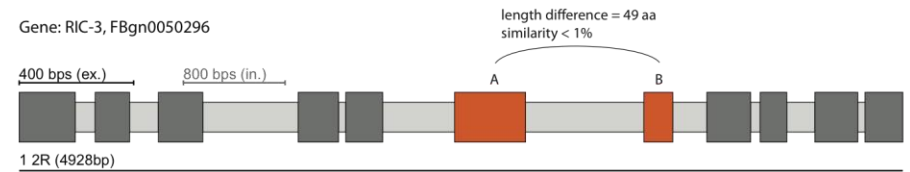
Gene: CG33012, FBgn0053012

According to RNASeq these exons are constitutive exons and are misannotated as MXE in *Dm* r5.36.



For clarity introns have been scaled up by a factor of 3.56

Gene: RIC-3, FBgn0050296



For clarity introns have been scaled down by a factor of 2.25

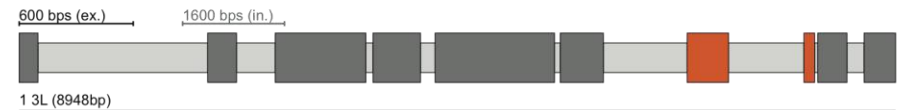
10 20 30 40 50 60 70 80

exonA GAATATAAAKKFAAKDTEKELYNASVSATEVASSLSASLQSHQQLKEAEQLMEIEKLRQKLESTERAMAQLVAEMFDTTAVST

exonB IVTAIQGLIDAADQLNGQDKQRATSDTETDSNK

Gene: CG14168, FBgn0036044

Both exons are not part of *Dm* r5.48 anymore.  
There is no RNASeq evidence for any of them.



For clarity introns have been scaled down by a factor of 2.99

Gene: TrpA1, Transient receptor potential A1, FBgn0035934



For clarity introns have been scaled down by a factor of 1.32

10 20 30

exonA IKYSFAFLQCFFMFAKIDEKTGESITTA SPIPLPALN

exonB IKYSFWFYQKTFEQIEA-KRKEFNDFKWRPAPLAVVN

10 20  
 .....|.....|.....|.....|.....  
 exonA 1 GLTNSPFRERVGSNRLPEKRPS  
 exonB 1 QHDTPRITEKHHNQLNSPLQS

For clarity introns have been scaled down by a factor of 26.36

```

      10      20      30      40      50      60      70      80
exonA  1  GYRLPVAAMFPLGAPVGAGAPTLTHLQLFVSMQMAQAPSAVAGGTAATSPATAAAAAAHAAAAAATFIMLAPRPHTAV
exonB  1  PAAAVAAAMRGVAIQRGHVGVGATPYHHFHHHPHLLAASAAAAQQQQQRQLAAAVATAAQAQQQQQQQAVVQQ
      90      100     110     120     130     140     150     160
exonA  81  QAAATPTAATPRRS
exonB  81  QQQVAAAQQHQHQHQHQHQHQVAVQQQQQVAVQQQQQHQHQHQHQHQHQHAAVAAAAAASHPHMHAHAHAHAHALGFLQAQ
      170     180     190     200     210     220     230     240
exonA
exonB  161 LQAVAVPTAASNAALQQSLAAIQNPSGNPNAAAAAAYAAARLSAATGATQSPQTAAAAAAAASMAASNAANANNAALH
      ...
exonA
exonB  241 GFAP

```

For clarity introns have been scaled down by a factor of 1.56

10 20 30 40  
 .....|.....|.....|.....|.....|.....|.....|.....|.....|.....  
 exonA 1 SRGHLASDLDPLGILTREKTVCKDGLARRANEDVLRQHSGLF  
 exonB 1 IRGHNIAHLDPLEINTP-----ELPGNSSTKSIYANFSF

For clarity introns have been scaled down by a factor of 1.81

For clarity introns have been scaled down by a factor of 1.53

10 20 30 40 50 60 70  
 exonA 1 TLFDADYFTEGRECVNGC-----AISTPLWRNDTGHYLCNACGLYMKMGMNRP LTKQPRRL  
 exonB 1 MAAESGGDFYKPNSEFNVGGGRSKANTSGAASSYSCPGSNATSAATS AVASGTAATAATTLDEHVSRRSRL

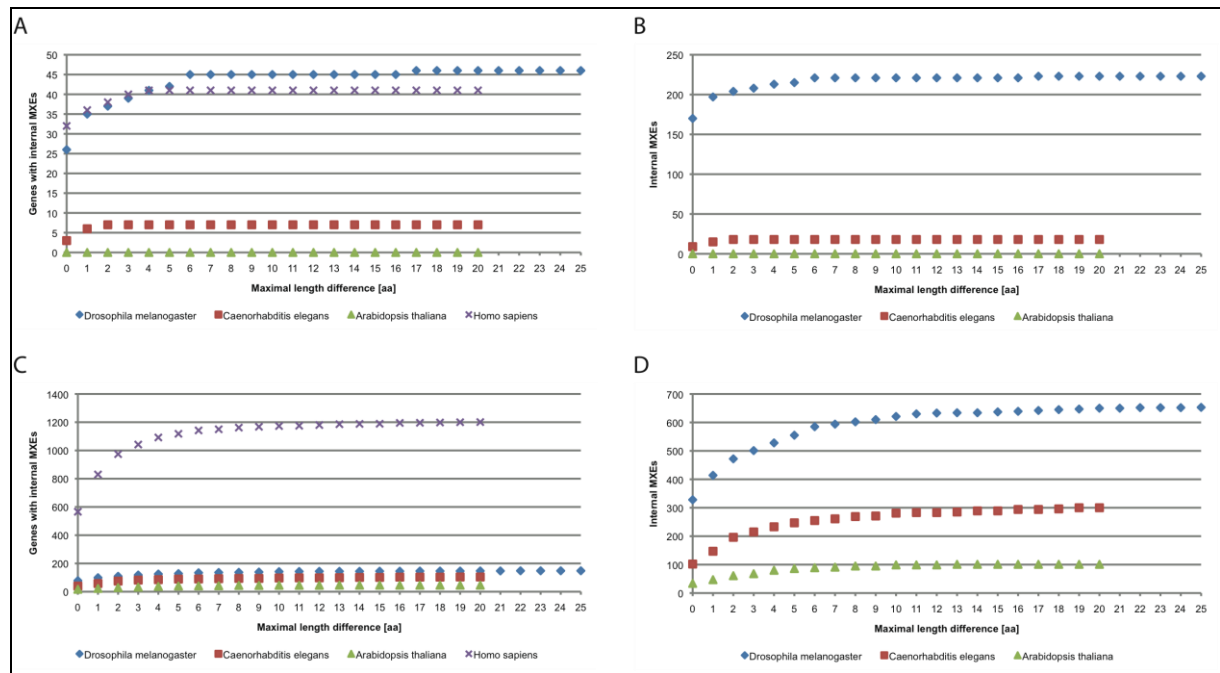
For clarity introns have been scaled down by a factor of 3.07

10 20  
 . . . . | . . . . | . . . . | . . . . | . . . . | . . . .  
 exonA 1 LSRLVSYDEQSQMTKPMMAVPQAKRGI  
 exonB 1 MERIFSGAWKEKHGEQEPEELATPTPLE

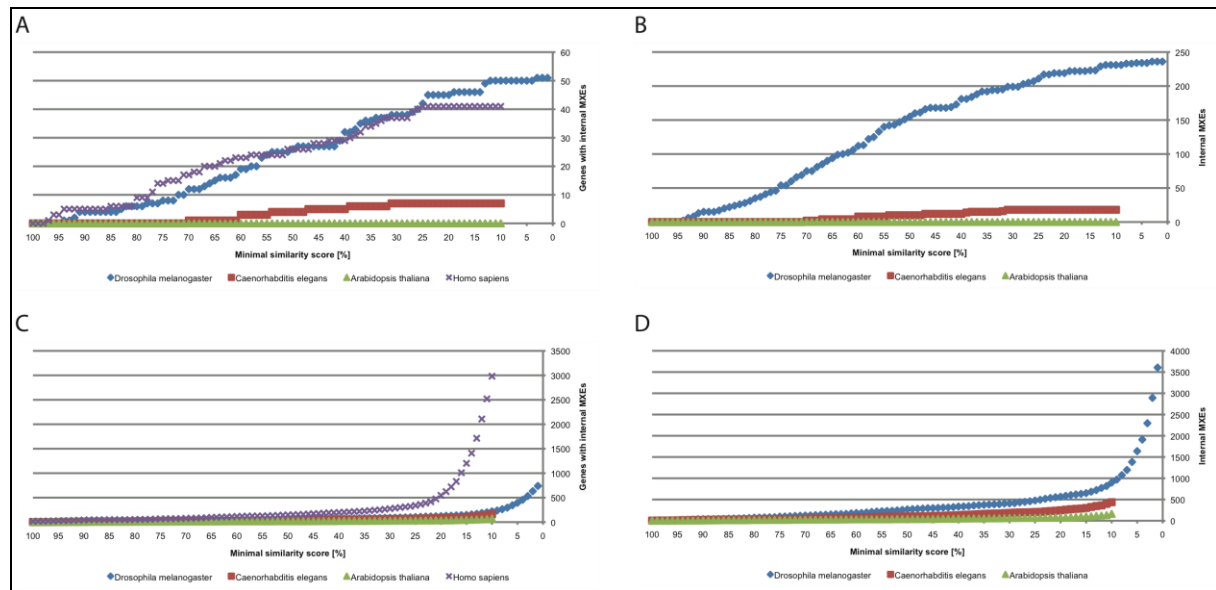
For clarity introns have been scaled down by a factor of 1.43

**Supplementary Figure S4. Genes containing annotated mutually exclusive spliced exons (MXEs), which could not be reconstructed using the default parameters.** These MXEs are shown in dark orange. MXEs found with the default prediction parameters are shown in light orange. Of

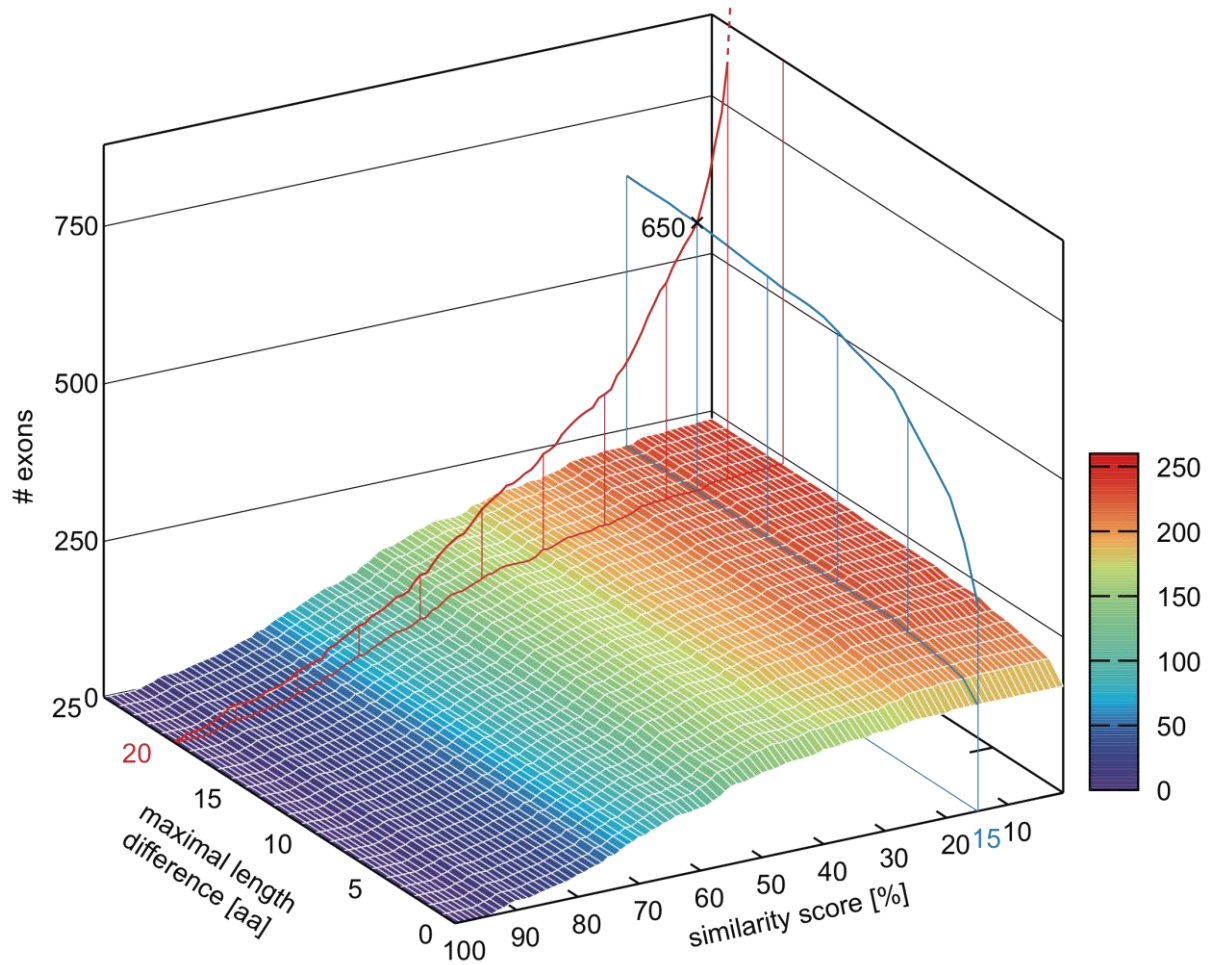
the annotated MXEs, which we could not reconstruct, four pairs of exons do not show any sequence similarity, three have length differences of more than 50 aa, three are annotated as differentially included in the latest release (Dm r5.48), one pair does not consist of neighboring exons, and two pairs of exons have completely been removed from the latest annotation. Thus, the sensitivity of our method is considerably higher than 83.5% (218 of the annotated internal MXEs reconstructed). All transcripts are represented 5' to 3'. The color coding is explained in the legend.



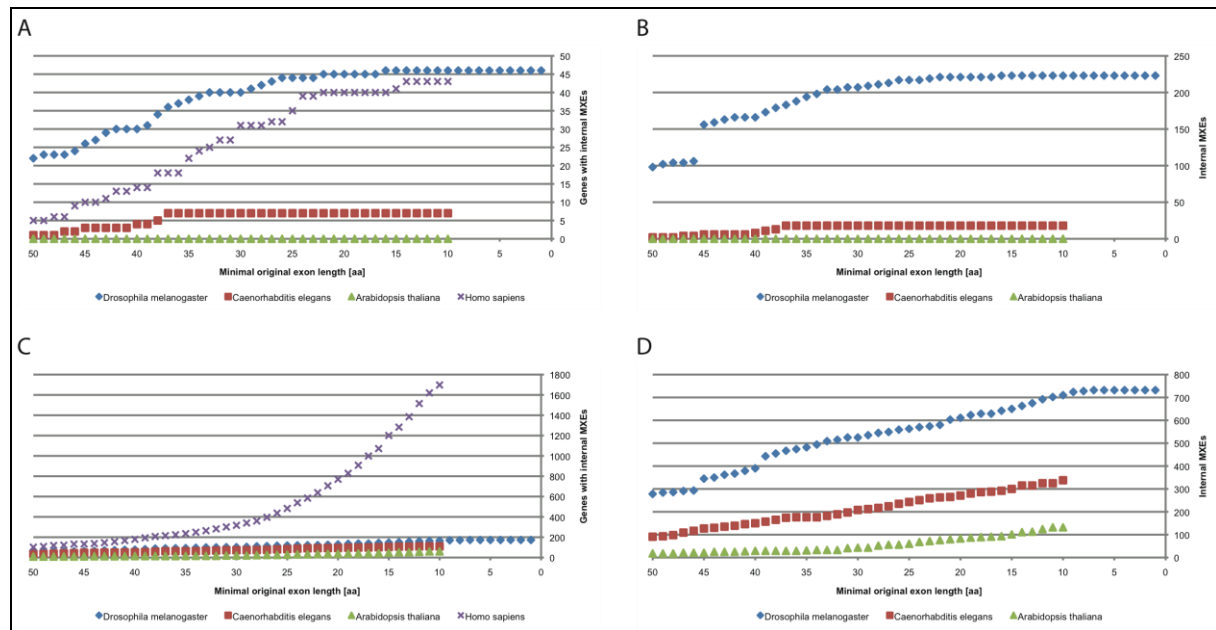
**Supplementary Figure S5. Reconstructed and predicted internal mutually exclusive spliced exons (MXEs) at a similarity score cut-off of 15%.** Apart from the MXEs that we cannot reconstruct because they are out of the scope of our preconditions (no sequence similarity, huge length difference), we assessed the sensitivity of our method when using a length difference of 20 residues and a similarity score of 15% as standard cut-offs. Given a similarity score of at least 15%, the analysis of the reconstructed MXEs shows that all annotated MXEs have length differences of less than 20 residues (A, B). A similar distribution is found for the length difference of the internal MXEs that we predict newly (C, D). A) Number of genes containing annotated internal MXEs that could be reconstructed at a given length difference cut-off having a similarity score of at least 15%. B) Number of annotated internal MXEs that could be reconstructed at a given length difference cut-off having a similarity score of at least 15%. C) Number of genes containing predicted internal MXEs (including annotated MXEs that could be reconstructed) with a similarity score of at least 15% at a given length difference. D) Number of internal MXE candidates (including annotated MXEs that could be reconstructed) with a similarity score of at least 15% at a given length difference.



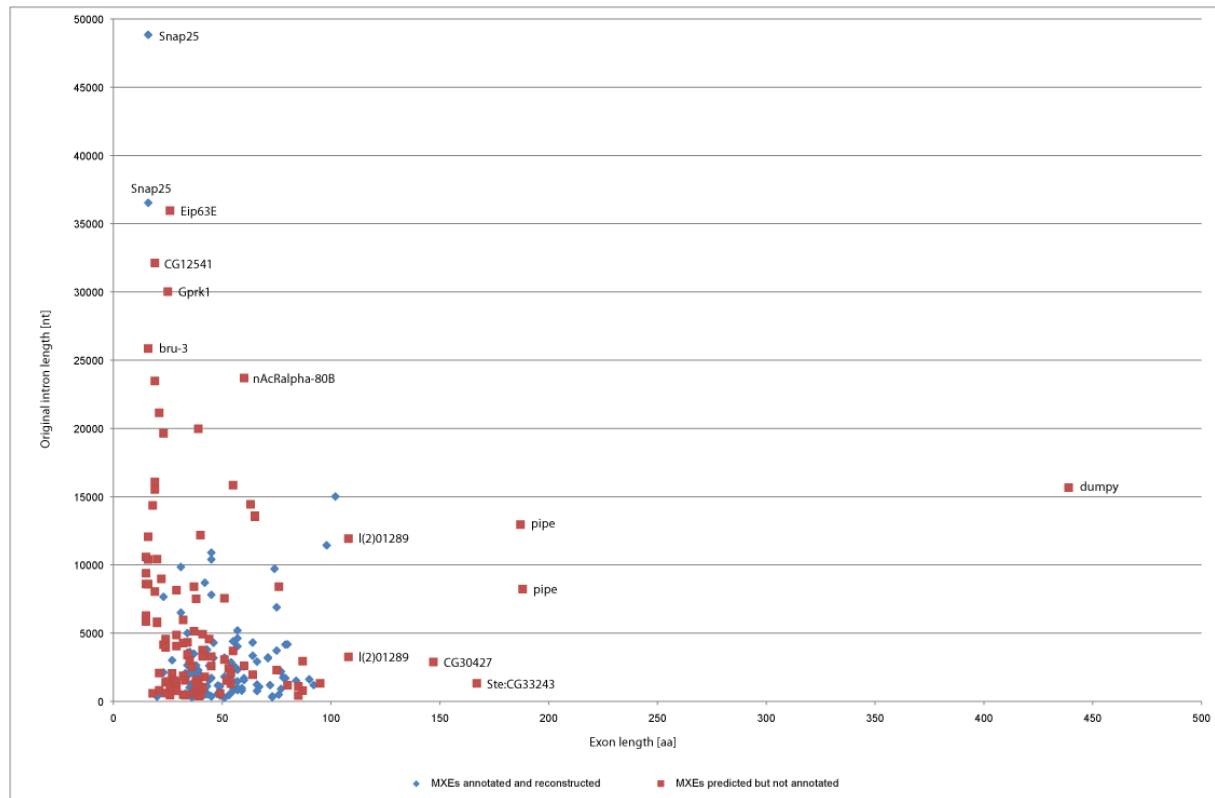
**Supplementary Figure S6. Reconstructed and predicted internal mutually exclusive spliced exons (MXEs) at a length difference cut-off of 20 residues.** To assess the suitability of the sequence similarity cut-off of 15% within the preconditions of our prediction method, we analysed the distribution of the annotated exons with a length difference of less than 20 residues (A, B). In contrast to the MXEs of *Hs* and *Ce*, the MXEs of *Dm* do not show a pronounced plateau. The number of predicted MXE candidates even shows an exponential increase below a similarity score of 10% (*Dm*) and 15% (*Hs*), respectively (C, D). A) Number of genes containing annotated internal MXEs that could be reconstructed at a given sequence similarity score cut-off and having a length difference of less than 20 aa. B) Number of internal MXEs that could be reconstructed at a given sequence similarity score cut-off and having a length difference of less than 20 aa. C) Number of genes containing internal MXE candidates (including annotated MXEs that could be reconstructed) predicted at a given sequence similarity score cut-off and having a length difference of less than 20 aa. D) Number of internal MXE candidates (including annotated MXEs that could be reconstructed) predicted at a given sequence similarity score cut-off and having a length difference of less than 20 aa.



**Supplementary Figure S7. Assessing annotated and predicted mutually exclusive spliced exons (MXEs) in *Drosophila melanogaster*.** This figure comprises information from the previous figures (Supplemental Figs. S4 and S5) for *Drosophila melanogaster* and shows the dependence of the number of internal MXEs on the maximal length difference and similarity between search exon and MXE candidate. The figure is similar to Fig. 1A of the main manuscript except that the number of exons is shown here in contrast to the number of genes, reflecting that many genes contain several clusters of MXEs and clusters with more than two MXEs. The colored grid denotes the number of MXEs as annotated in FlyBase r5.36 that were also predicted by WebScipio. The red and blue lines mark the number of predicted MXE candidates at the maximal length difference of 20 amino acids and at the minimal similarity score of 15%, respectively.

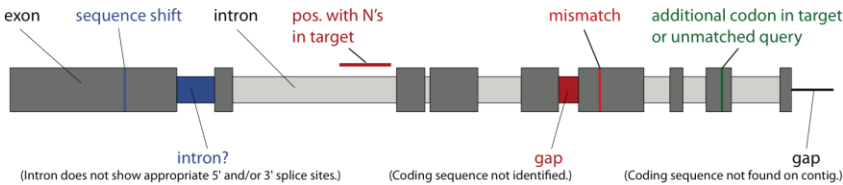


**Supplementary Figure S8. Reconstructed and predicted internal mutually exclusive spliced exons (MXEs) in dependence of a minimal original exon length.** The sequences of very short exons do not contain enough complexity to exclude the identification of “similar” exons, especially if they are surrounded by long introns. Luckily, short exons within genes are rather rare and are predominantly found at gene borders. In order to avoid the inclusion of many false positives we introduced the parameter “minimal original exon length”. Annotated MXEs, which we can reconstruct with a length difference cut-off of 20 residues and a similarity score cut-off of 15%, are all longer than ten residues (A, B). For the initial search for MXE candidates in *Drosophila* we set this parameter to one residue (A, B). However, only a few candidates were found for exons shorter than 15 residues. Therefore, we set the minimal original exon length parameter to 15 residues for the analysis of the *Drosophila* genome and for the search for MXE candidates in the other model organisms (C, D). The value seems appropriate for *Caenorhabditis* and *Arabidopsis* while the number of MXE candidates is increasing exponentially in dependence of the search exon length in human. This is most probably due to the much longer introns in human compared to the other species analysed. A) Number of genes containing annotated internal MXEs in dependency of the length of the MXEs that could be reconstructed at a sequence similarity score cut-off of 15% and a length difference of less than 20 aa. B) Number of annotated internal MXEs in dependency of the length of the MXEs that could be reconstructed at a sequence similarity score cut-off of 15% and a length difference of less than 20 aa. C) Number of genes containing internal MXE candidates in dependency of the length of the MXEs that were predicted at a sequence similarity score cut-off of 15% and a length difference of less than 20 aa. D) Number of internal MXE candidates in dependency of the length of the MXEs that were predicted at a sequence similarity score cut-off of 15% and a length difference of less than 20 aa.



**Supplementary Figure S9. Scatter plot of the internal mutually exclusive spliced exon (MXE) candidates.** Blue, annotated in r5.36; red, predicted MXEs. This figure is similar to Fig. 1B of the main manuscript. However, some of the genes containing either very long introns or very long exons, for which MXE candidates were predicted, are indicated. If exons are short the complexity of the translations will be low and chances will thus be high to predict false positive candidates, especially if the surrounding introns are long. The introns surrounding annotated MXEs vary from 50 to 50,000 nucleotides. Although most introns range up to 15,000 nucleotides we therefore cannot assume that potential MXE candidates in longer introns are false predictions. MXE candidates, which are also conserved in other arthropods, were found for example in very long introns of the *nAcRalpha-80B* and *bruno-3* genes. In the case of long exons, it is very unlikely that by chance the translation of intronic region shows sequence similarity to neighbouring exons. However, if long exon candidates are found in long introns these could also, instead of being part of a cluster of MXEs, belong for example to mis-annotated tandemly arrayed gene duplicates or belong to the very rare cases of clusters of exons, which share sequence homology and are spliced as cluster. Here, we also found false positive MXE candidates, that are annotated in the latest FlyBase release as belonging to different tandemly arrayed gene duplicates (*CG33243* gene region; FlyBase r5.48), that were derived from isoforms containing different, mutually exclusive clusters of exons (*CG30427* gene; *pipe* gene<sup>49</sup>) and that were part of some isoforms of the gigantic *dumpy* gene that displays a complex pattern of alternative splicing<sup>50</sup>.

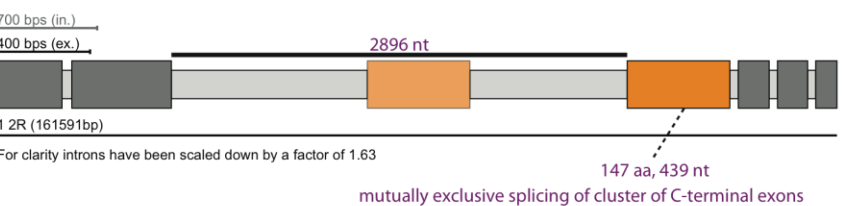
Legend



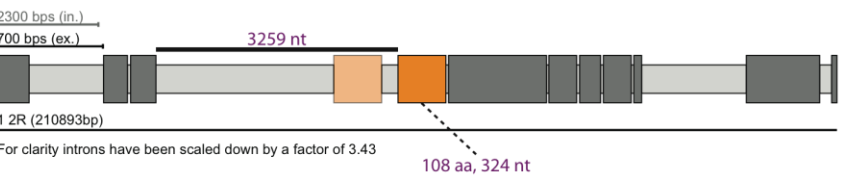
Gene: dp, dumpy, FBgn0053196  
Polypeptide: FBpp0293673



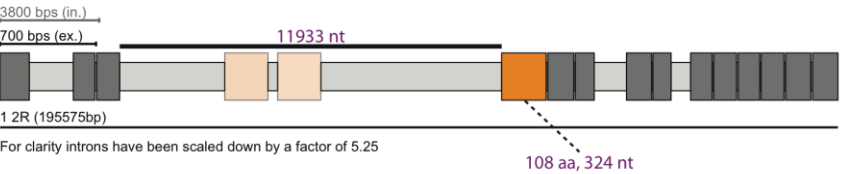
Gene: CG30427, FBgn0043792  
Polypeptide: CG30427-PB, FBpp0072320



Gene: l(2)01289, lethal (2) 01289, FBgn0010482  
Polypeptide: l(2)01289-PA, FBpp0085469



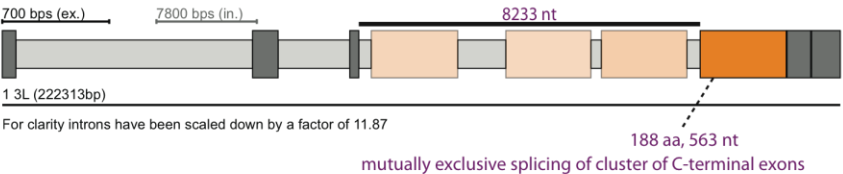
Gene: l(2)01289, lethal (2) 01289, FBgn0010482  
Polypeptide: l(2)01289-PH, FBpp0290635



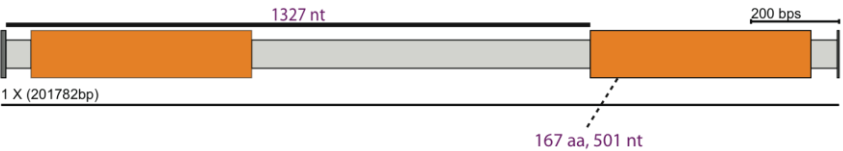
Gene: pip, pipe, FBgn0003089  
Polypeptide: pip-PA, FBpp0074777



Gene: pip, pipe, FBgn0003089  
Polypeptide: pip-PG, FBpp0074775

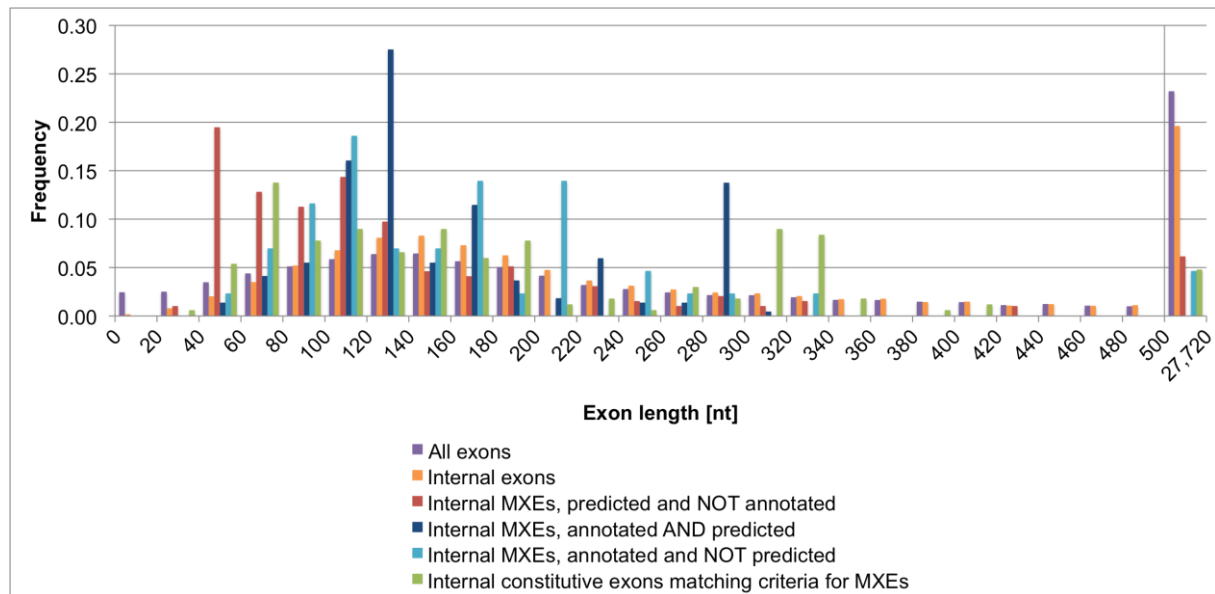


Gene: Ste:CG33243, FBgn0053243  
Polypeptide: Ste:CG33243-PB, FBpp0289369

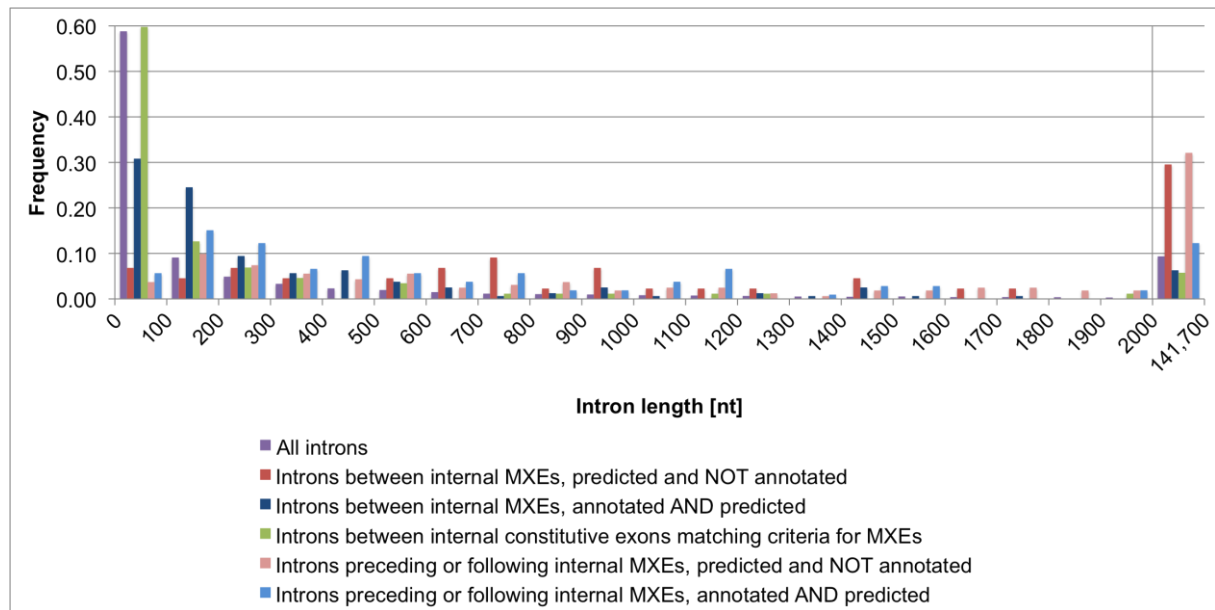


**Supplementary Figure S10. Examples of mutually exclusive spliced exon (MXE) candidates found for long introns.**

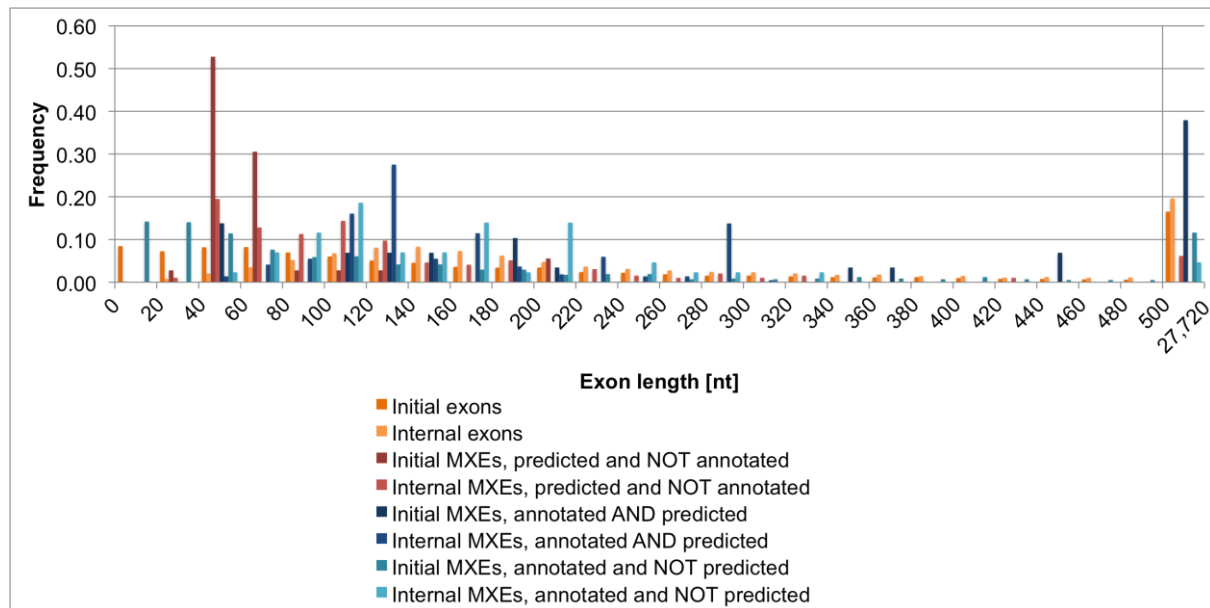
All transcripts are represented 5' to 3'. Colored big bars represent MXEs. The darkest colored bar is the exon that was included in the query sequence, while the lighter colored bars represent identified MXEs. The higher the similarity between the candidate and the query exon the darker the color of the candidate (100% identity would result in the same color). The opacity of the colors of each alternative exon corresponds to the alignment score of the alternative exon to the original one. The color coding is explained in the legend.



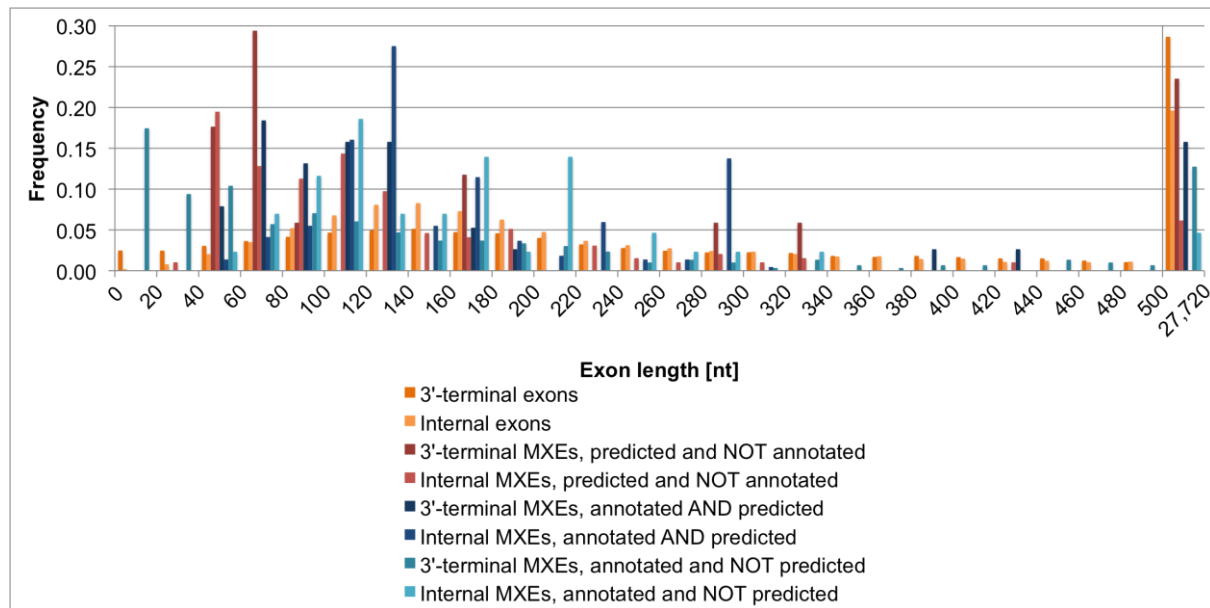
**Supplementary Figure S11. Comparison of exon lengths.** Various subsets of annotated and predicted mutually exclusive spliced exons (MXEs) are compared to all exons and internal constitutive exons sharing our criteria for MXEs. The exon lengths of the annotated and predicted MXEs show almost the same distribution like all exons of *Drosophila* with a broad peak around 140 residues. Interestingly, there is a second smaller peak for the length of MXEs at 300 amino acids. The comparison of the annotated MXEs to the predicted MXE candidates shows similar distributions meaning that the predictions represent normal MXEs. The internal MXEs that are annotated and that we cannot reconstruct also display a similar distribution but in addition tend to represent larger exons as compared to the other sets. Surprisingly, the constitutive exons sharing our criteria for MXEs show three striking peaks at 80, 320 and 340 residues but show a local minimum at 140 residues. This supports the notion that the predicted MXEs rather represent MXEs than potential constitutively spliced exons.



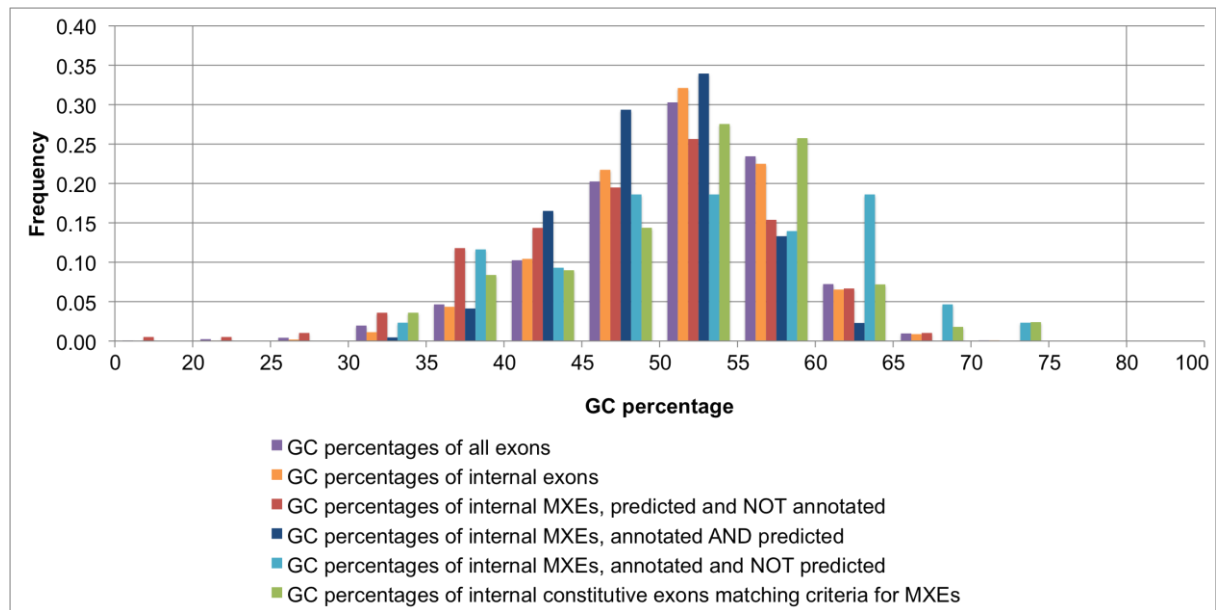
**Supplementary Figure S12. Comparison of intron lengths.** Introns next to various subsets of annotated and predicted mutually exclusive spliced exons (MXEs) are compared to all introns and introns next to internal constitutive exons sharing our criteria for MXEs. The comparison of the intron lengths shows a broad distribution with a tendency to rather short introns (< 300 bp).



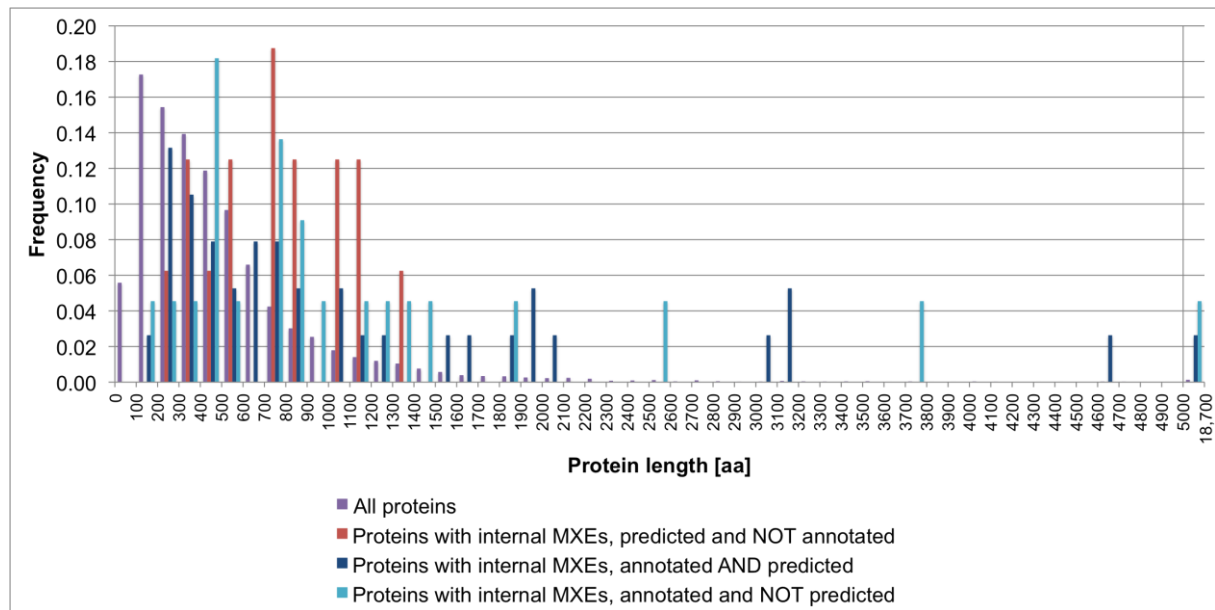
**Supplementary Figure S13. Comparison of exon lengths of initial exons of multi-exon genes.** Various subsets of annotated and predicted initial exons matching the criteria for mutually exclusive spliced exons (MXEs) are compared to all exons and internal MXEs. Because the algorithm is based on protein coding sequence it could be possible that the initial and terminal exons of the coding region are not the initial and terminal exons of the transcripts. In this case, these exons would be regarded as internal exons. Therefore, we also analysed candidate exons of initial and terminal exons that share the criteria of MXEs. In general, initial and terminal exons of multi-exon genes are considerably shorter than internal exons. Some of these match the criteria of MXEs. Of those, almost all code for at least 40 residues. In these cases it is unlikely that pseudo-duplicates of low-complexity exons were found.



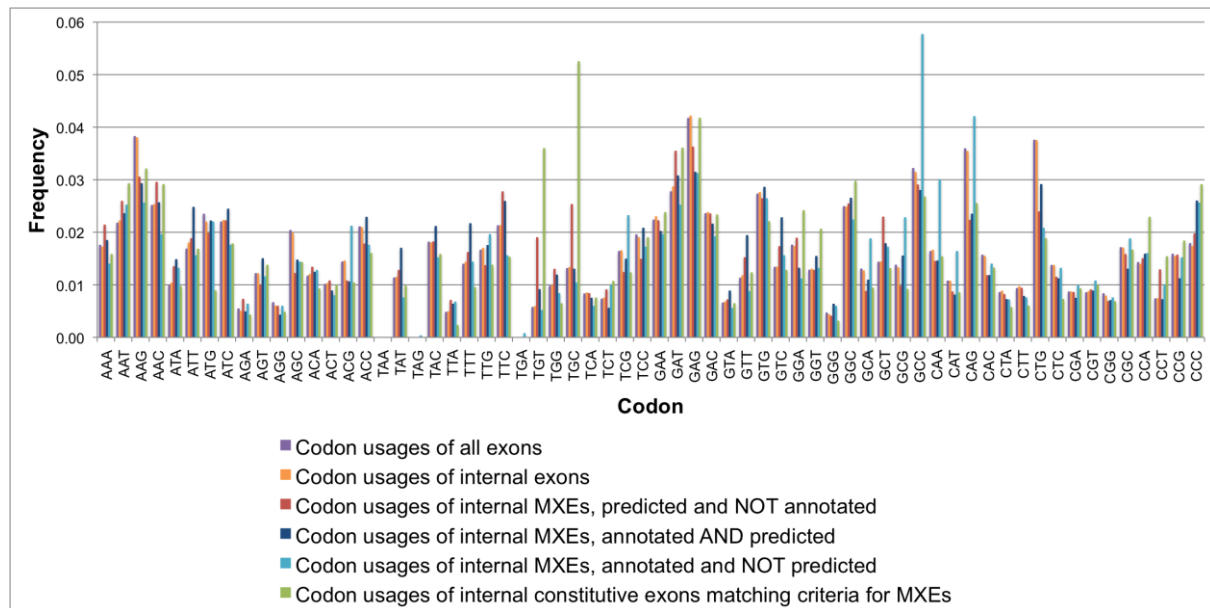
**Supplementary Figure S14. Comparison of exon lengths of terminal exons of multi-exon genes.** Various subsets of annotated and predicted terminal exons matching the criteria for mutually exclusive spliced exons (MXEs) are compared to all exons and internal MXEs. Because the algorithm is based on protein coding sequence it could be possible that the initial and terminal exons of the coding region are not the initial and terminal exons of the transcripts. In this case, these exons would be regarded as internal exons. Therefore, we also analysed candidate exons of initial and terminal exons that share the criteria of MXEs. In general, initial and terminal exons of multi-exon genes are considerably shorter than internal exons. Some of these match the criteria of MXEs. Of those, almost all code for at least 40 residues. In these cases it is unlikely that pseudo-duplicates of low-complexity exons were found.



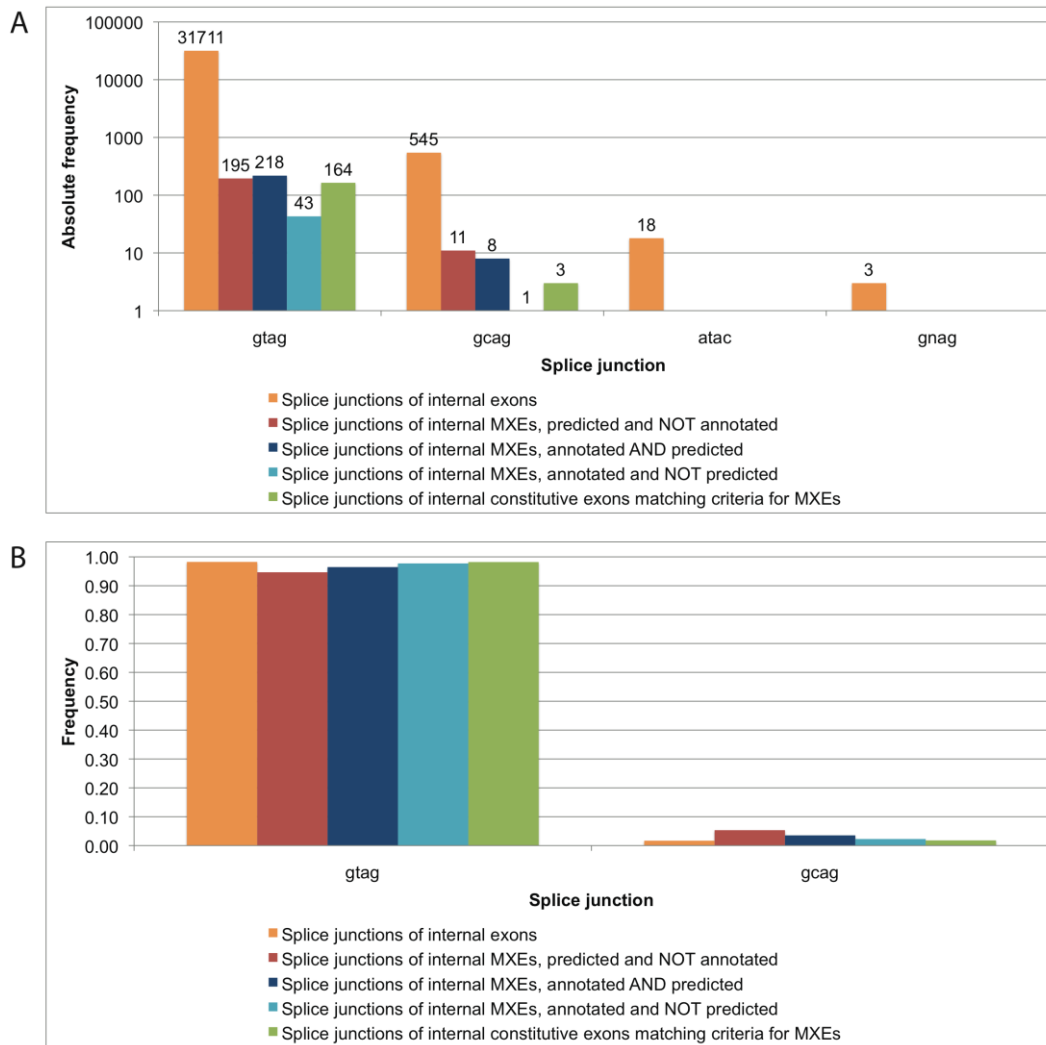
**Supplementary Figure S15. Comparison of GC content of exons.** The GC content of all exons (reference) is compared to the GC content of annotated and predicted internal mutually exclusive spliced exons (MXEs) and to internal constitutive exons sharing our criteria for MXEs. The GC content of all exons shows a broad distribution around 55%. The MXEs, which we cannot reconstruct, and the constitutive exons sharing our criteria of MXEs have a broader GC content distribution with a remarkably higher percentage of exons with GC contents of 60 to 75%. The distribution of the GC content of the predicted MXEs is similar to the distribution of the annotated MXEs except for a slight increase of exons with GC contents of 40 to 45%.



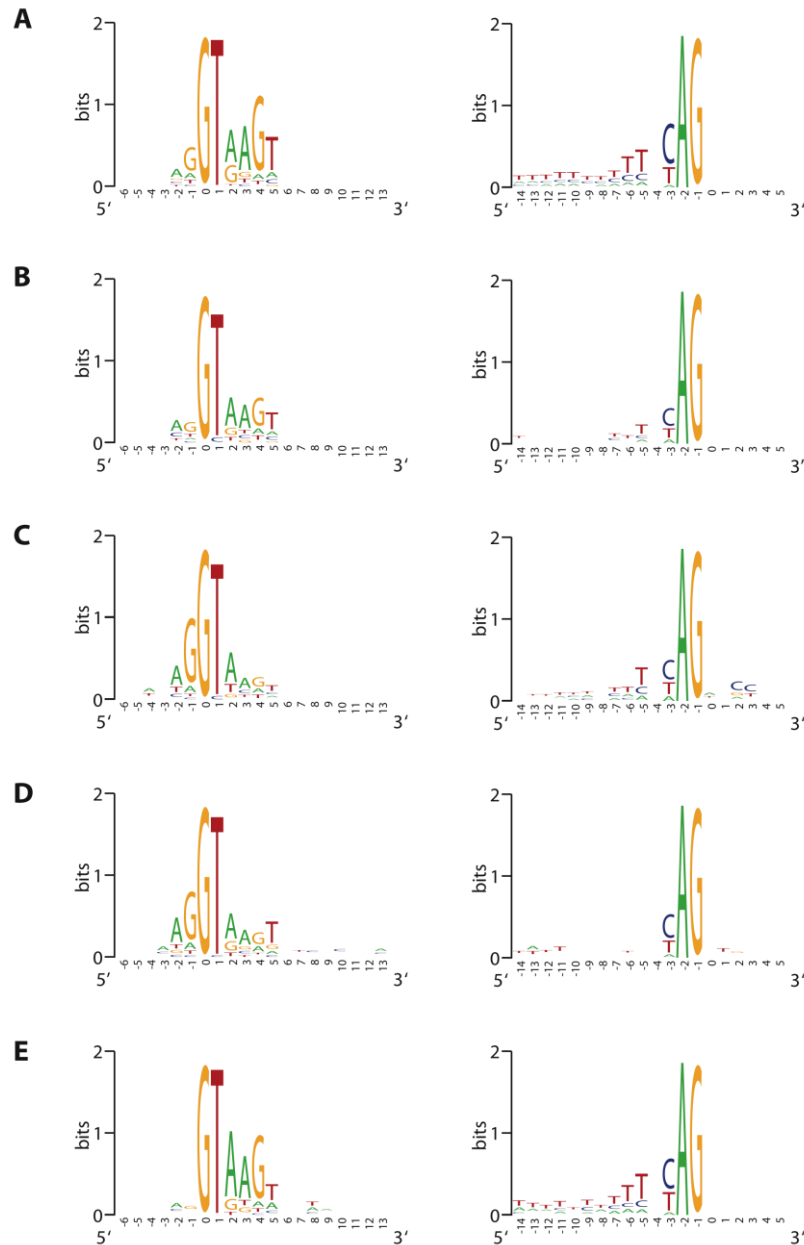
**Supplementary Figure S16. Comparison of the lengths of the translations of one isoform per gene.** For the reconstruction of the translations of the genes containing mutually exclusive spliced exons (MXEs) only one isoform has been chosen and only one exon of each cluster. For the protein lengths of all proteins, only the isoforms “A” were considered. To assess whether MXEs are predominantly found in proteins of a certain size, we analysed the lengths of the translations. Here, from each alternatively spliced gene (independently of alternative splicing type) only one transcript and the corresponding translation were considered. Proteins built with MXEs are relatively longer than the average proteins. The distribution of the proteins with annotated MXEs and with predicted MXE candidates is very similar.



**Supplementary Figure S17. Comparison of the codon usage.** Codon usage in all exons is compared to that of genes containing annotated or predicted mutually exclusive spliced exons (MXEs) and to that of internal constitutive exons sharing our criteria for MXEs. The codon usage of the MXEs (annotated and predicted) is very similar to the codon usage of all or all internal exons except for the codons AAG, AGC CAG and CTG that are slightly less represented in MXEs. Strikingly, the percentage of cysteine-coding codons (TGT and TGC) is five times higher in constitutive exons sharing our criteria of MXEs compared to all exons, and the MXEs, that are annotated in FlyBase but that we cannot reconstruct, have a considerably higher content of alanines (GCC codon) and glutamines (CAA and CAG codons).

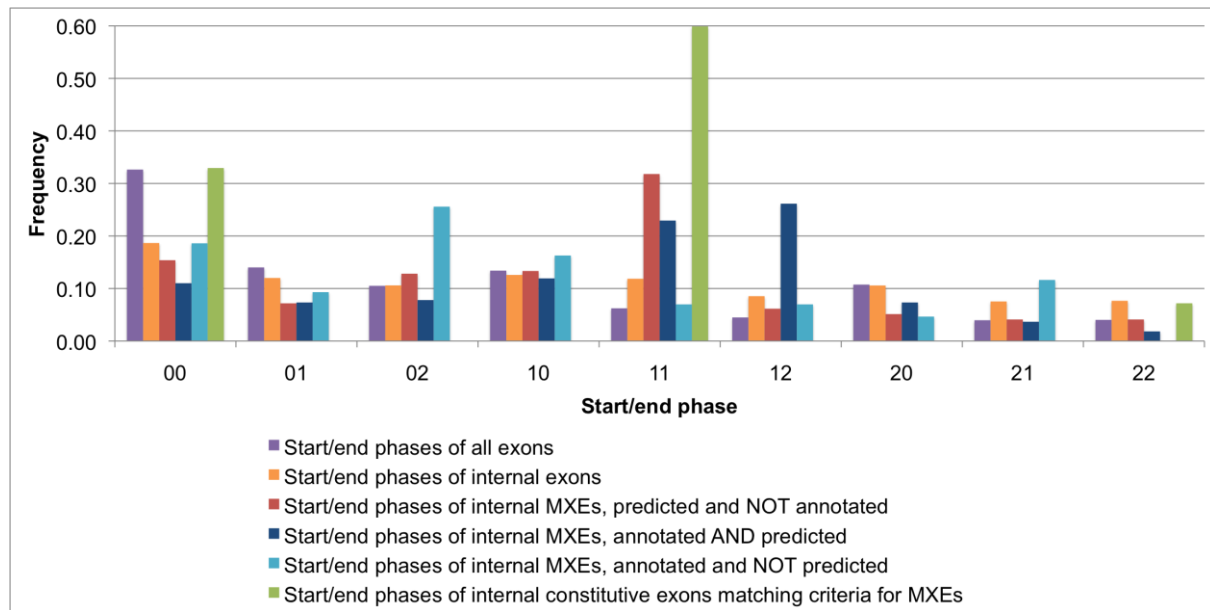


**Supplementary Figure S18. Comparison of splice junctions.** The splice junctions of all introns are compared to those of the putative introns between a mutually exclusive spliced exon (MXE) and the next constitutive exon before and after a cluster of MXEs. MXEs are separated in annotated or predicted MXEs and compared to internal constitutive exons sharing our criteria for MXEs. Total numbers are given in (A) and percentages in (B). As known, by far most introns have the splice junctions GT---AG followed by the GC---AG splice junctions (A). Only a few of the annotated introns have other splice junctions. The percentage of the GC---AG splice junction in introns surrounding MXEs is slightly higher than that of all introns (B). These numbers are, however, hard to interpret because the total number of MXEs spliced by GC---AG is very low.



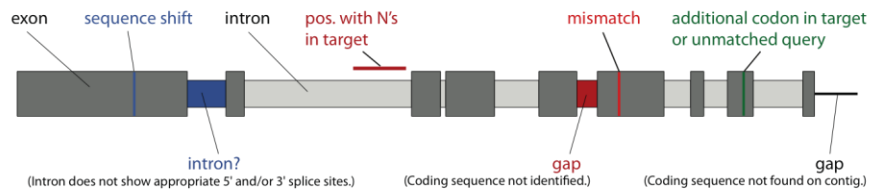
**Supplementary Figure S19. Conservation of intron splice junctions.** The weblogos were generated from the aligned 14 nucleotides of the intron and six nucleotides of the exon of both the 5'- and 3'-splice sites. The height of the letters represents the degree of conservation. A) All internal introns. B) Predicted internal mutually exclusive spliced exons (MXEs) that were not annotated. C) Annotated and reconstructed internal MXEs. D) Annotated but not reconstructed internal MXEs. E) Internal constitutive exons matching our criteria of MXEs. Splice junctions display sequence conservation beyond the two-base splice site.

Characteristic to all internal exons (pattern strongly dominated by constitutive exons) and the constitutive exons sharing our criteria of MXEs are the considerably stronger conservation of the bases AGT in positions +4, +5 and +6 of the intron. In contrast, the introns following the MXEs (annotated and predicted) have a stronger conserved G in position -1. The 3' ends of the introns before the MXEs have similar patterns as compared to all introns.

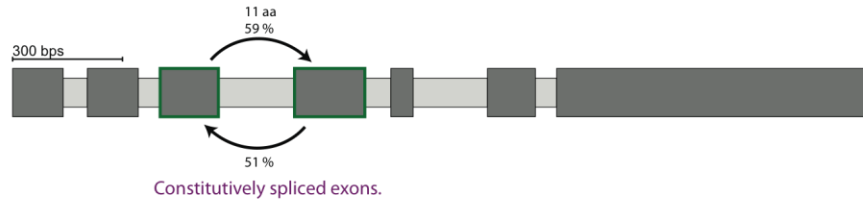


**Supplementary Figure S20. Comparison of start/end phases of exons.** A strong indication for mutually exclusive splicing is the impossibility to incorporate more than one of the mutually exclusive spliced exons (MXEs) of a cluster into the final transcript because of the incompatibility of the splice site phases. Exons can be classified based on the phase of the flanking intron: symmetric exons are 0-0 (intron interrupts the reading frame between two consecutive codons), 1-1 (intron interrupts the reading frame between the first and second base of a codon) and 2-2, and asymmetric exons are 0-1, 0-2, 1-0, 1-2, etc. Symmetric exons are the only ones that can be spliced in succession without changing the reading frame. Thus, constitutive exons sharing our criteria of MXEs comprise only symmetric exons. Compared to the annotated MXEs, the predicted MXEs show a slightly higher percentage of symmetric exons. Therefore, these potential exon candidates could also be spliced constitutively or they could be incorporated in a differentially included manner.

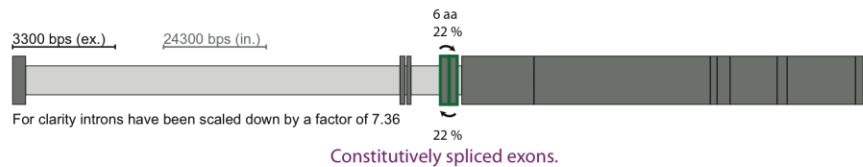
## Legend



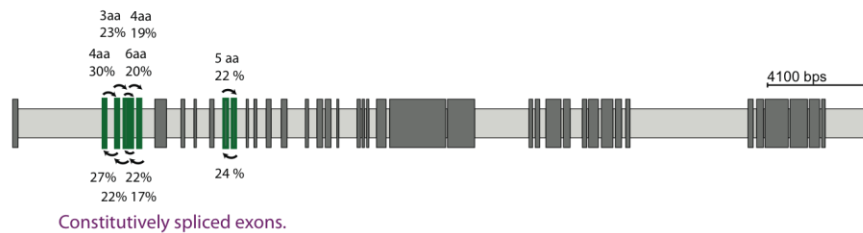
Gene: C901, FBgn0021742  
Polypeptide: C901-PA, FBpp0073256



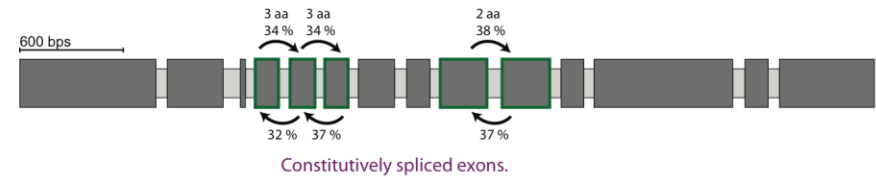
Gene: Megalin, FBgn0261260  
Polypeptide: Megalin-PA, FBpp0291363



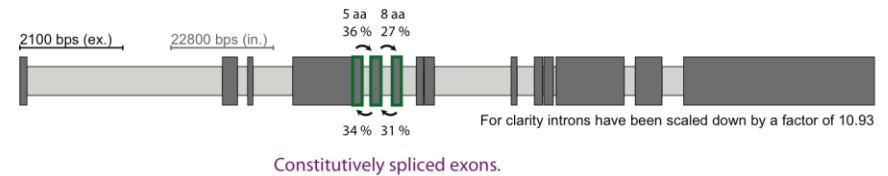
Gene: trol, terribly reduced optic lobes, FBgn0261451  
Polypeptide: trol-PD, FBpp0070440



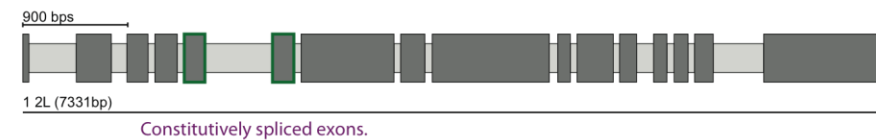
Gene: CG15570, FBgn0029697  
Polypeptide: CG15570-PA, FBpp0070613



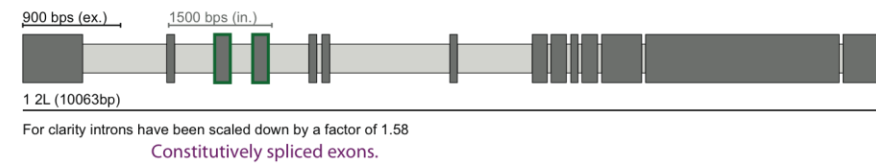
Gene: Ten-a, Tenascin accessory, FBgn0259240  
Polypeptide: Ten-a-PD, FBpp0289136



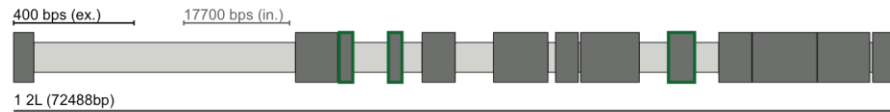
Gene: CG10186, FBgn0032797  
Polypeptide: CG10186-PA, FBpp0080818



Gene: rk, rickets, FBgn0003255  
Polypeptide: rk-PA, FBpp0080183



Gene: nAcRalpha-30D, nicotinic Acetylcholine Receptor  $\alpha$  30D, FBgn0032151  
 Polypeptide: nAcRa-30D-PD, FBpp0079503

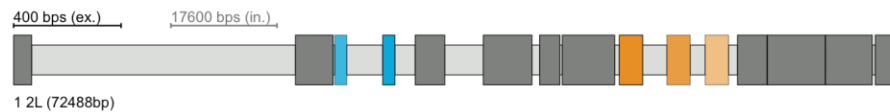


For clarity introns have been scaled down by a factor of 50.34

Polypeptide: nAcRa-30D-PE, FBpp0079502



For clarity introns have been scaled down by a factor of 48.97



For clarity introns have been scaled down by a factor of 44.59

RNASeq supports mutually exclusive splicing

RNASeq supports differentially included splicing, last exon candidate not reported before.

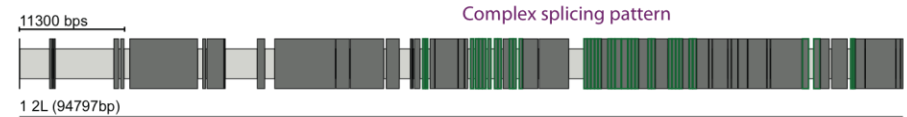
Gene: CG8086, FBgn0032010  
 Polypeptide: CG8086-PG, FBpp0297483



Constitutively spliced.

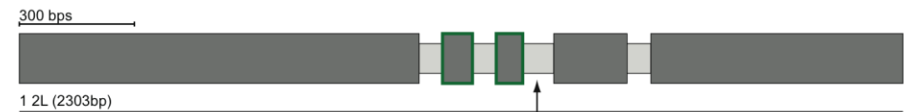
Differentially included splicing supported by many cDNAs.

Gene: dp, dumpy, FBgn0053196  
 Polypeptide: FBpp0288445



Complex splicing pattern

Gene: Rrp1, Recombination repair protein 1, FBgn0004584  
 Polypeptide: Rrp1-PA, FBpp0077362



Intron retention

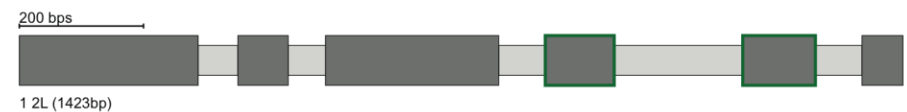
Gene: CG10039, FBgn0031581  
 Polypeptide: CG10039-PA, FBpp0077196



For clarity introns have been scaled down by a factor of 3.71

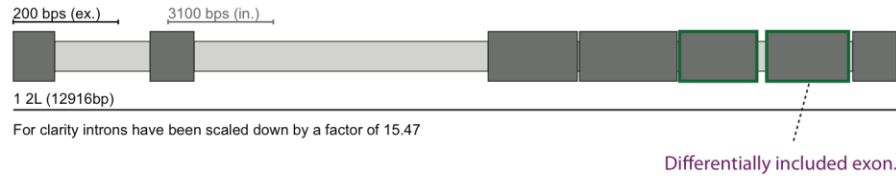
Two separate genes. Tandem gene duplicates. Annotation corrected in FlyBase.

Gene: Tsp26A, Tetraspanin 26A, FBgn0031760  
 Polypeptide: Tsp26A-PB, FBpp0111937

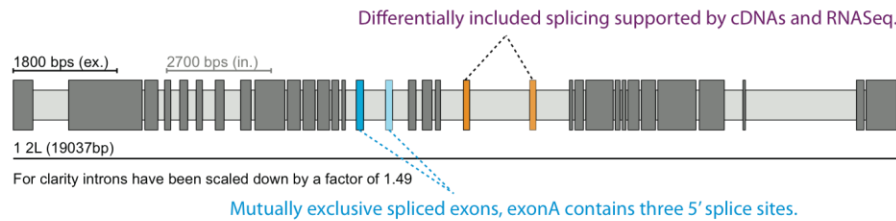
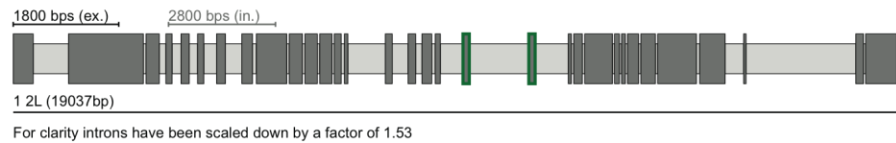


Differentially included splicing supported by cDNAs and RNASeq.

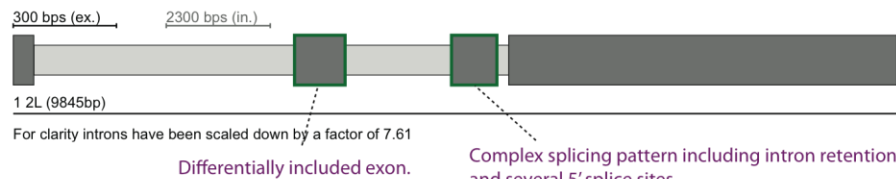
Gene: stai, stathmin, FBgn0051641  
Polypeptide: stai-PB, FBpp0078828



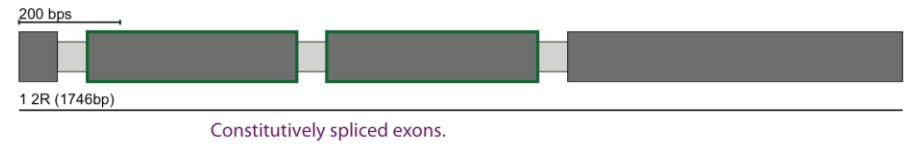
Gene: Ca-alpha1D, Ca<sup>2+</sup>-channel protein  $\alpha$ 1 subunit D, FBgn0001991  
Polypeptide: Ca-alpha1D-PC, FBpp0089047



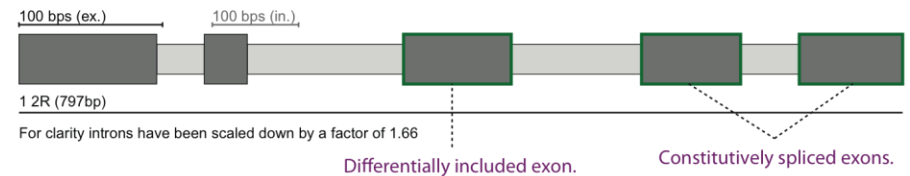
Gene: CG5674, FBgn0032656  
Polypeptide: CG5674-PA, FBpp0080574



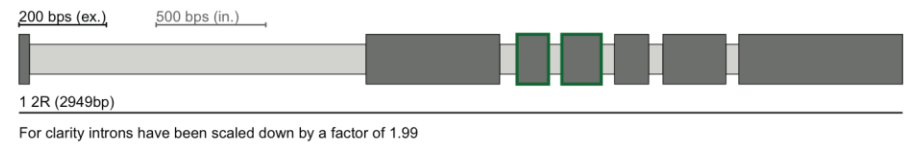
Gene: CG10494, FBgn0034634  
Polypeptide: CG10494-PA, CG10494-PA



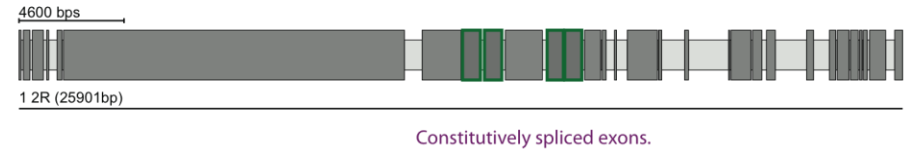
Gene: CG13428, FBgn0034515  
Polypeptide: CG13428-PA, FBpp0085579



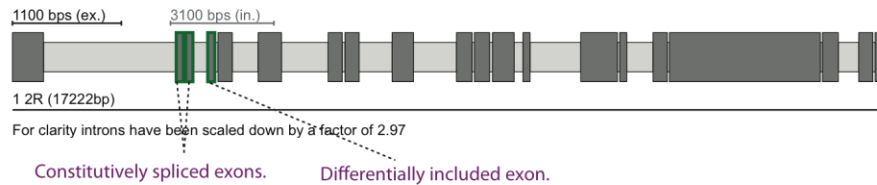
Gene: CG15615, FBgn0034159  
Polypeptide: CG15615-PB, FBpp0289779



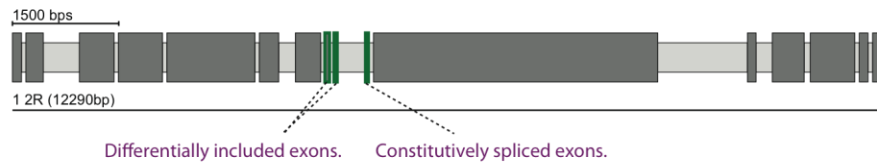
Gene: Strn-Mlck, Stretchin-Mlck, FBgn0013988  
Polypeptide: Strn-Mlck-PD, FBpp0086409



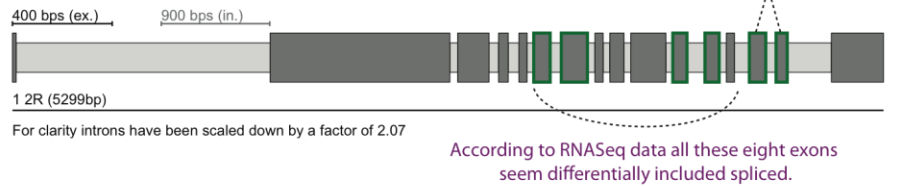
Gene: sli, slit, FBgn0003425  
Polypeptide: sli-PC, FBpp0086438



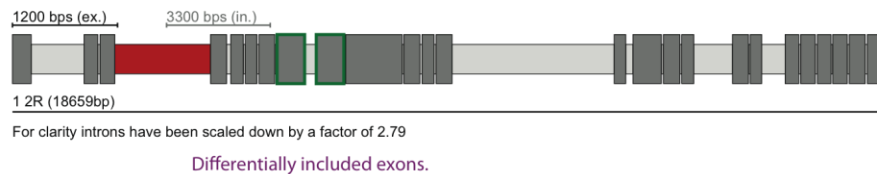
Gene: tou, toutatis, FBgn0033636  
Polypeptide: tou-PA, FBpp0087193



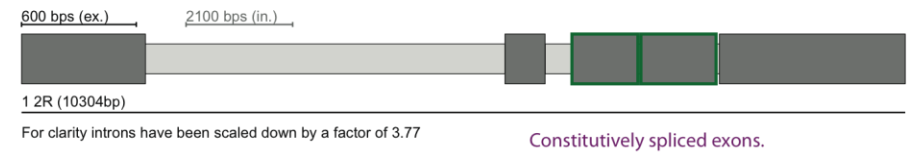
Gene: Cpr47Ef, Cuticular protein 47Ef, FBgn0033603  
Polypeptide: Cpr47Ef-PD, FBpp0291859



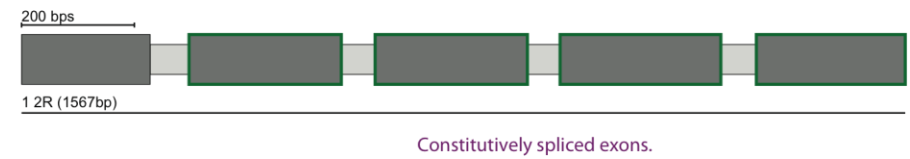
Gene: l(2)01289, lethal (2) 01289, FBgn0010482  
Polypeptide: l(2)01289-PB, FBpp0085470



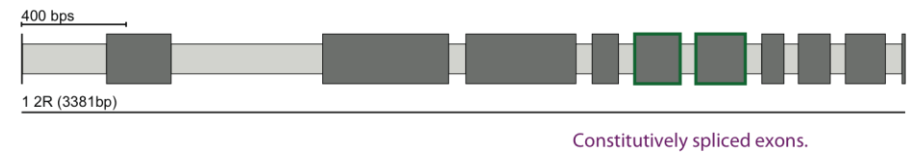
Gene: rgr, regular, FBgn0033310  
Polypeptide: rgr-PA, FBpp0087772



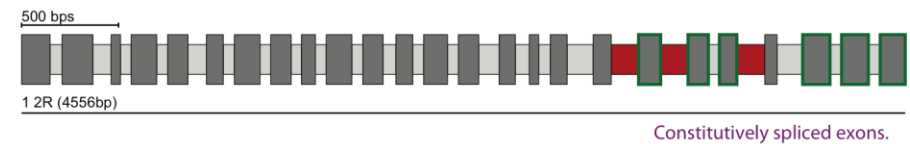
Gene: CG6357, FBgn0033875  
Polypeptide: CG6357-PA, FBpp0086764



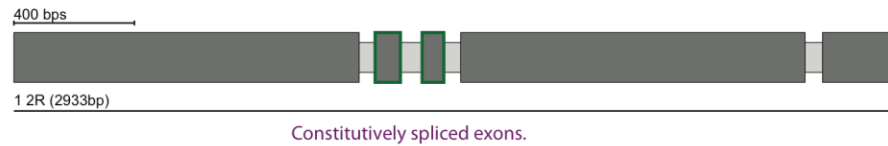
Gene: Dek, FBgn0026533  
Polypeptide: Dek-PA, FBpp0099855



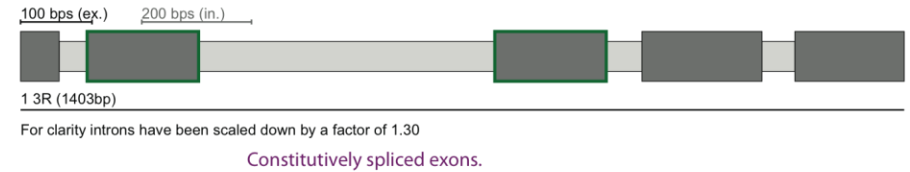
Gene: CG30395, FBgn0050395  
Polypeptide: CG30395-PB, FBpp0289463



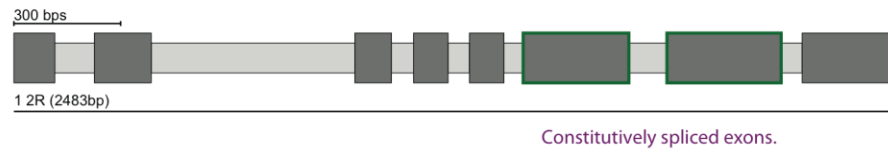
Gene: CG9861, FBgn0034844  
 Polypeptide: CG9861-PA, FBpp0071911



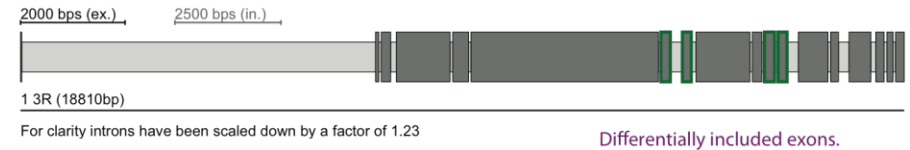
Gene: CG33483, FBgn0053483  
 Polypeptide: CG33483-PB, FBpp0292484



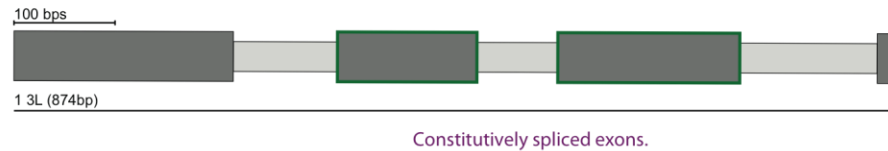
Gene: Mlp60A, Muscle LIM protein at 60A, FBgn0259209  
 Polypeptide: Mlp60A-PB, FBpp0288975



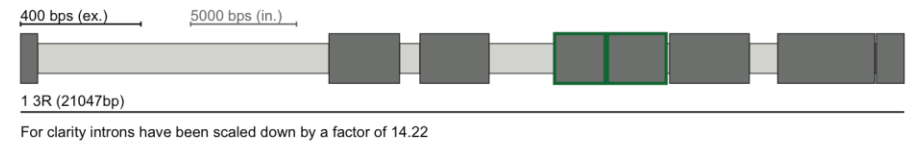
Gene: Ppn, Papilin, FBgn0003137  
 Polypeptide: Ppn-PE, FBpp0291051



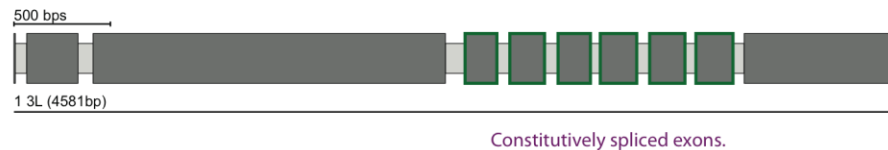
Gene: miple, FBgn0027111  
 Polypeptide: miple-PA, FBpp0072405



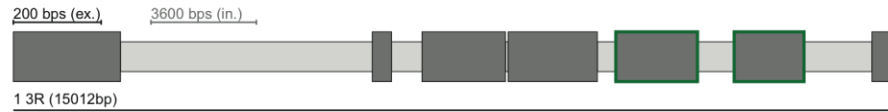
Gene: betaTub97EF,  $\beta$ -Tubulin at 97EF, FBgn0003890  
 Polypeptide:  $\beta$ Tub97EF-PA, FBpp0084630



Gene: CG6947, FBgn0036233  
 Polypeptide: CG6947-PA, FBpp0075777



Gene: tau, FBgn0051057  
 Polypeptide: tau-PA, FBpp0084567



For clarity introns have been scaled down by a factor of 15.01

Differentially included exons.

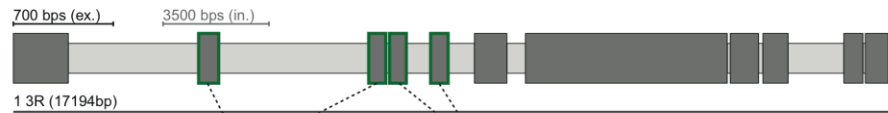
Gene: CG31406, FBgn0051406  
 Polypeptide: CG31406-PA, FBpp0081713



For clarity introns have been scaled down by a factor of 1.63

Constitutively spliced exons.

Gene: LpR1, Lipophorin receptor 1, FBgn0066101  
 Polypeptide: LpR1-PK, FBpp0290685



For clarity introns have been scaled down by a factor of 4.72

Constitutively spliced exons. Differentially included exons.

Gene: CG9297, FBgn0038181  
 Polypeptide: CG9297-PA, FBpp0082295



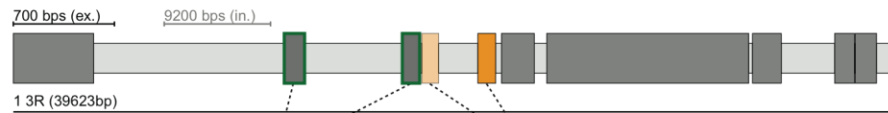
For clarity introns have been scaled down by a factor of 1.24

Constitutively spliced exons.

Gene: LpR2, Lipophorin receptor 2, FBgn0051092  
 Polypeptide: LpR2-PA, FBpp0084301



For clarity introns have been scaled down by a factor of 13.16



For clarity introns have been scaled down by a factor of 12.50

Constitutively spliced exons. Mutually exclusive exons

Gene: CG42342, FBgn0259244  
 Polypeptide: CG42342-PD, FBpp0289172



For clarity introns have been scaled down by a factor of 23.57

Constitutively spliced exons.

Gene: Fsh, Fsh-Tsh-like receptor, FBgn0016650  
 Polypeptide: Fsh-PA, FBpp0082933



Constitutively spliced exons.

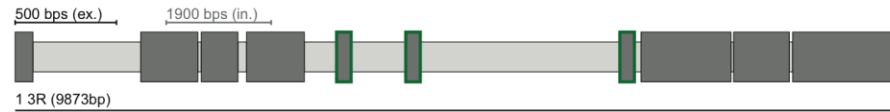
Gene: CG5621, FBgn0038840  
 Polypeptide: CG5621-PB, FBpp0110256



For clarity introns have been scaled down by a factor of 1.18

Differentially included exons according to RNASeq data.

Gene: Lgr3, FBgn0039354  
 Polypeptide: Lgr3-PA, FBpp0084273



For clarity introns have been scaled down by a factor of 3.67

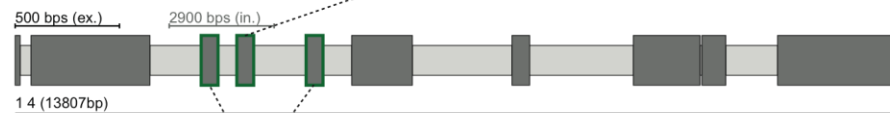
Constitutively spliced exons.

Gene: CG9682, FBgn0039760  
 Polypeptide: CG9682-PA, FBpp0084981



Constitutively spliced exons.

Gene: CG1674, FBgn0039897  
 Polypeptide: CG1674-PB, FBpp0088185



For clarity introns have been scaled down by a factor of 5.77

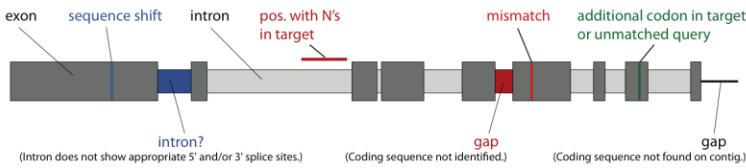
Differentially included exon.

Constitutively spliced exons.

**Supplementary Figure S21. List of genes containing constitutive exons matching the prediction parameters for mutually exclusive spliced exons (MXEs).** Several of these exons are even annotated as MXEs in the latest Flybase release on RNA-Seq evidence, including a cluster of MXEs

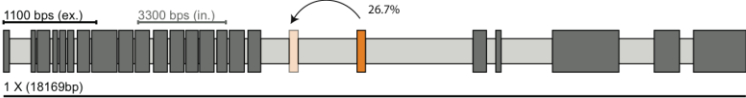
in the  $\beta$ Tub97EF gene, the Lipophorin receptor 1 gene, and the nicotinic Acetylcholine Receptor  $\alpha$  30 D gene. Another gene is now split into two tandemly arrayed duplicates (CG10039 is now CG43773 and CG43774). The putative constitutive exons in 15 other genes are now annotated as differentially included or as other types of alternative splice forms. All transcripts are represented 5' to 3'. The color coding is explained in the legend. Colored big bars represent MXEs. The darkest colored bar is the exon that was included in the query sequence, while the lighter colored bars represent identified MXEs. The higher the similarity between the candidate and the query exon the darker the color of the candidate (100% identity would result in the same color). The opacity of the colors of each alternative exon corresponds to the alignment score of the alternative exon to the original one. The green strokes mark constitutive exons that match our criteria for MXEs.

Legend



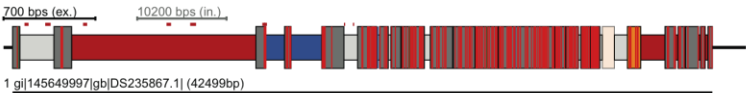
Gene: mmd, mind-meld, FBgn0259110  
Polypeptide: mmd-PD, FBpp0288810

Exon A is annotated in r5.48.  
Supported by RNA-Seq data.  
Conserved in Anopheles gambiae, Pediculus humanus corporis, dana, dere, dgri, dmoj, dper, dsec, dsim, dvir, dwil and dyak.  
RNA-Seq: Exon B has an alternative splice site at the 3'-end.



For clarity introns have been scaled down by a factor of 3.17

Cross-species search in Pediculus humanus corporis

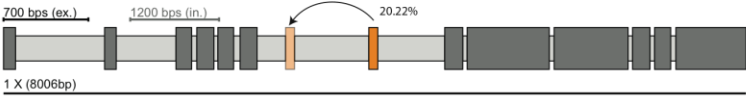


For clarity introns have been scaled down by a factor of 15.00

```
.....10.....20.....30.....
exonA  GTSDQNRISTLTMTVILLTVIVKCVFISFATLAVCYR-
exonB  --ENYHGSNTVFLVGVLMSSVVGVFITFTLMALCYR-
AngExonA --DHDQGVSTLAMVIMLVIVKCVFLCFALMAVCYR-
AngExonB --ANYHGSNTVFLVGVLMSSVVGVFITFTLMALCYRS
PdcExonA --GGNNNNLSTLAMVFILVGVGVFCFTLMAVCYR-
PdcExonB --ENYHSTNTGFLVGVLMSSVVGVFILFALMALCYR-
```

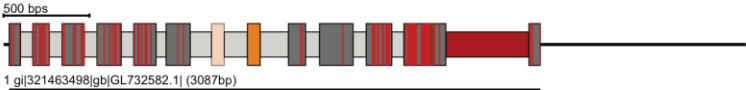
Gene: g, garnet, FBgn0001087  
Polypeptide: g-PB, FBpp0073673

Exon A is annotated in r5.48.  
Supported by EST and RNA-Seq data.  
Conserved in Aedes aegypti, Anopheles gambiae, Apis mellifera, Atta cephalotes, Daphnia pulex, dana, dere, dgri, dmoj, dper, dpse, dsec, dvir, dwil and dyak.  
RNA-Seq: Exon A has an alternative splice site at the 5'-end.



For clarity introns have been scaled down by a factor of 1.64

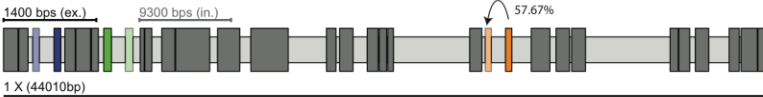
Cross-species search in Daphnia pulex



```
.....10.....20.....
exonA  FASLTTFEPALGRKLTQPLIEIIS
exonB  FGALTPLEPRLGKKLIEPLTNLIHS
DapExonA LNAMTQCDRLSKCISQPLIAIIS
DapExonB FGALTPLEPRLGKKLIEPLTNLIHS
```

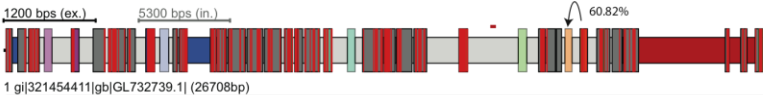
Gene: cac, cacophony, FBgn0263111  
Polypeptide: cac-PA, FBpp0298319

Exon A is annotated in r5.48.  
Supported by RNA-Seq data.  
Conserved in Aedes aegypti, Anopheles gambiae, Apis mellifera, Atta cephalotes, Daphnia pulex, Pediculus humanus corporis, dana, dere, dgri, dmoj, dper, dpse, dsec, dvir, dwil and dyak.



For clarity introns have been scaled down by a factor of 6.73

Cross-species search in Daphnia pulex

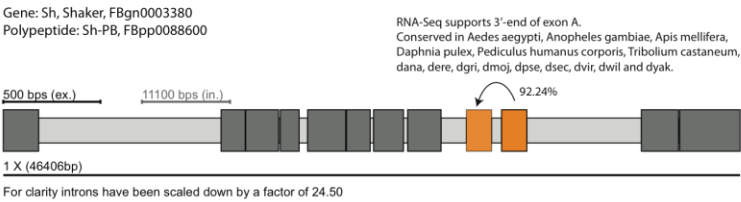
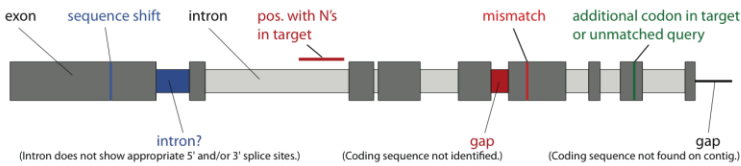


For clarity introns have been scaled down by a factor of 4.46

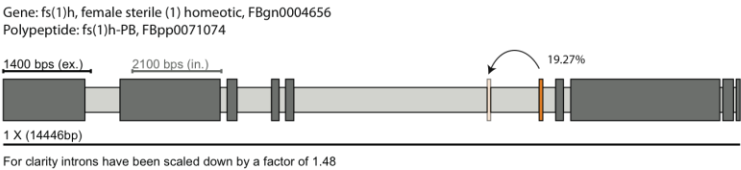
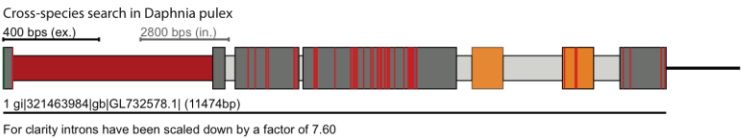
```
.....10.....20.....30.....
exonA  VFGNIRYDF-DTQLNRHNNFQSFSGGIMLLFR
exonB  VFGNIKLTGVENSITRHHNNFQSFQGVMLLFR
DapExonA VFGNHLHDF-DSSVNRHNNFQSFQGLLLLFR
DapExonB VFGNILLEFGTTHIHRHNNFRSFIQGLMLLFR
```



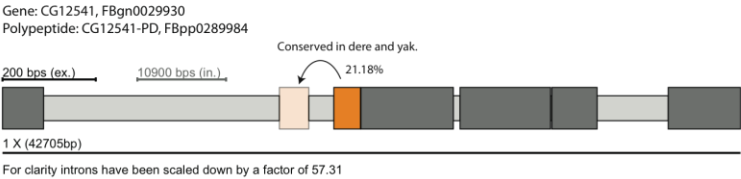
Legend



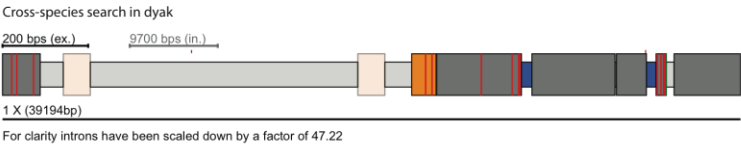
```
.....10.....20.....30.....40.....
exonA  GVVLFSSAVYFAEAGSDSSFFKSI PDGFVAVVIMTTVGYGDMR
exonB  GVVLFSSAVYFAEAGSENSFFKSI PDGFVAVVIMTTVGYGDMT
DapExonA GVVLFSSAVYFAEAGSEYSEKSI PDGFVAVVIMTTVGYGDMR
DapExonB GVVLFSSAVYFAEAGSEVSEKSI PDGFVAVVIMTTVGYGDMT
```



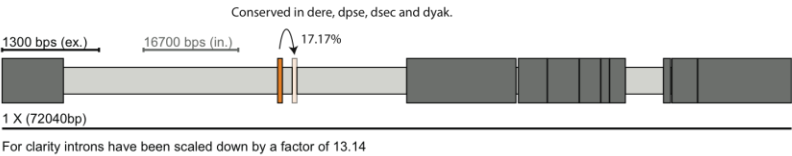
```
.....10.....20.....
exonA  GRGNKKKGRKKSGR-RELRN
exonB  GDGDERPPRRKKSRDSNGSN
```



```
.....10.....20.....
exonA  VRFMRSLMIAERASTKASLKY
exonB  V--VRLEVF AEVTTAASLSE
dyakExonA VIVAHTFAFEICVVTLAMCSS
dyakExonB VRFMGSQVFAVRLSAKASLKY
dyakExonC V--VRLEVF AEELTTAAALSE
```



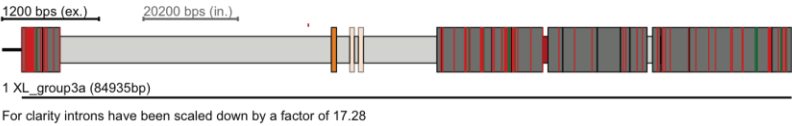
Gene (r5.36): CG42248  
Polypeptide(r5.36): CG42248-PD, FBpp0288785  
Gene (r5.48): CG43867, FBgn0264449  
Polypeptide (r5.48): CG43867-PD, FBpp0304858



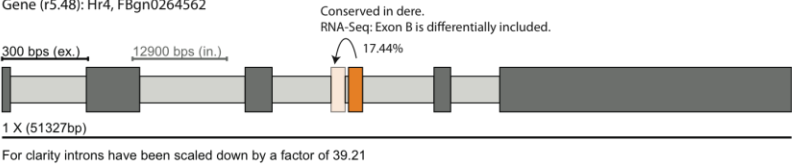
10 20

exonA L T E L E Q R V I E A E R A E E A E D K  
exonB A S T W Q L A V L E S V E N A G K S A R K  
dpseExonA L T E L E Q R V I E A E R A E E A E D K  
dpseExonB L R G I E R N - - T A R E R E S D V E E R  
dpseExonC A T A R E Q R S C A A C E R E S A A R T C

Cross-species search in dpse



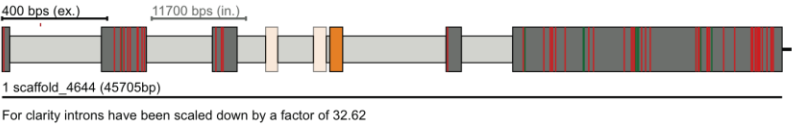
Gene (r5.36): CG3600  
Polypeptide(r5.36): CG3600-PC, FBpp0288868  
Gene (r5.48): Hr4, FBgn0264562



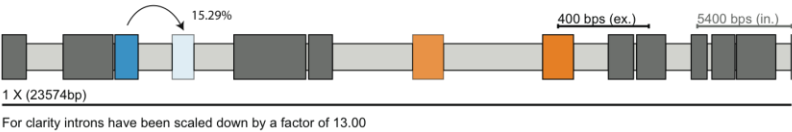
10

exonA R A R S A V G Q R F V G G R F I  
exonB T C Q A E E G Q S S A G S H Y T  
dereExonA R C - H E L G E R S S T S T W N  
dereExonB S E R S A V G Q R F V G G R F I  
dereExonC T C Q A E E G Q S S A G S H Y T

Cross-species search in dpse



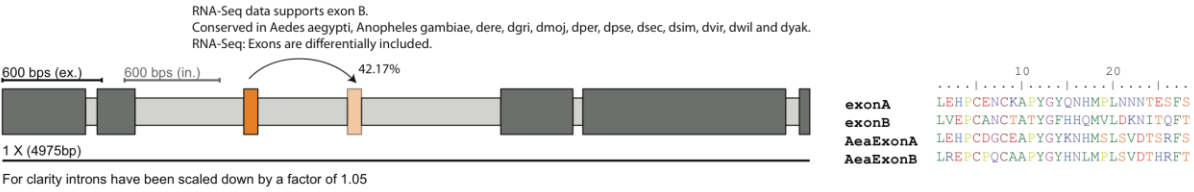
Gene: SK, small conductance calcium-activated potassium channel, FBgn0029761  
Polypeptide: SK-PH, FBpp0289694



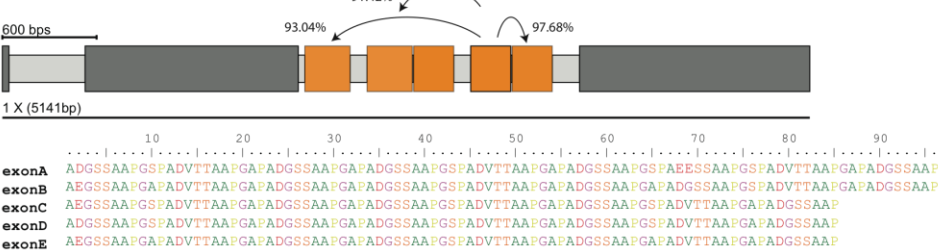
10 20 30

exonA A S F Y S T A L K T L I S V S T V I L L G L I V A Y H A L E V Q V R  
exonB K S H N S Y S L H T I C S L S L S I I - - I I F N Q C L P P Q I N

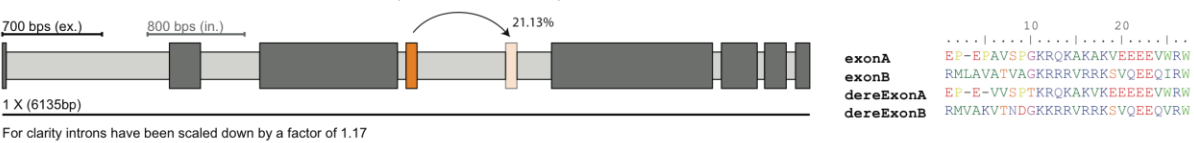
Gene: mys, mysospheroid, FBgn0004657  
Polypeptide: mys-PA, FBpp0071061



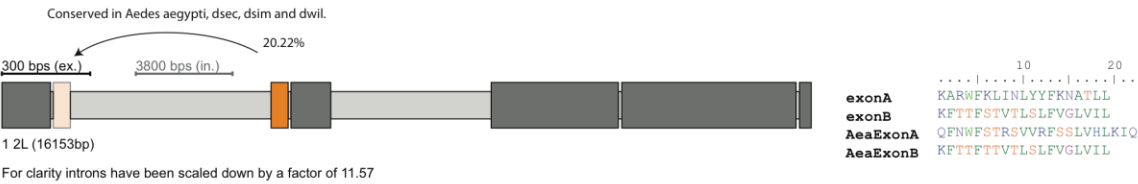
Gene: Muc11A, Mucin 11A, FBgn0052656  
Polypeptide: Muc11A-PA, FBpp0088744



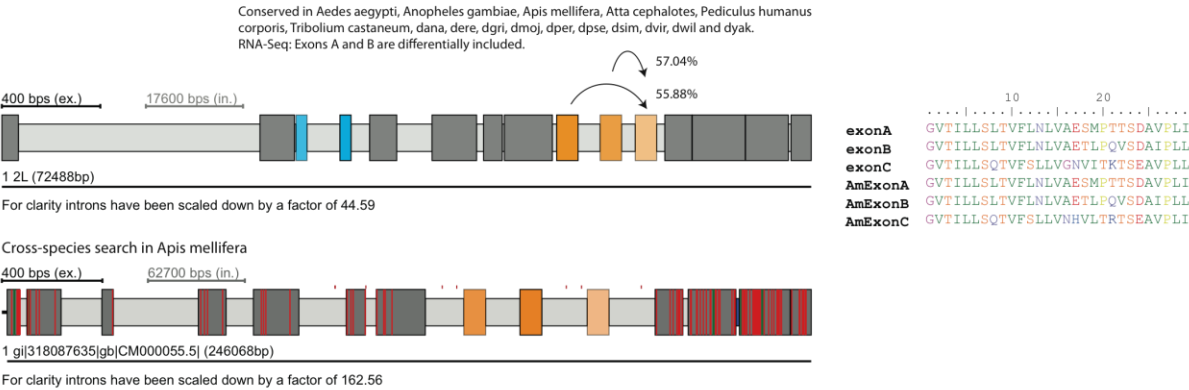
Gene: Top1, Topoisomerase 1, FBgn0004924  
Polypeptide: Top1-PA, FBpp0073822



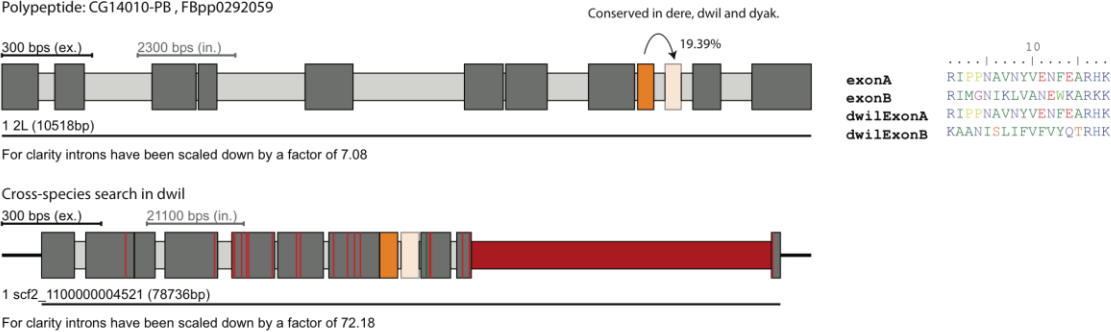
Gene: mol, moladietz, FBgn0086711  
Polypeptide: mol-PA, FBpp0080238



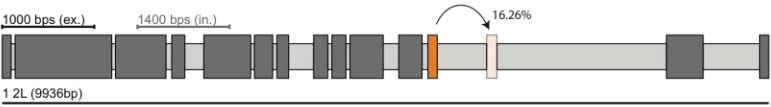
Gene: nAcRalpha-30D, nicotinic acetylcholine receptor alpha 30D, FBgn0032151



Gene: CG14010, FBgn0031725  
Polypeptide: CG14010-PB, FBpp0292059



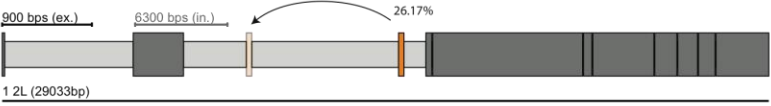
Gene: tim, timeless, FBgn0014396  
Polypeptide: tim-PB, FBpp0077256



For clarity introns have been scaled down by a factor of 1.40

```
.....10.....20.....30.....
exonA Y--TPDPTP-FVPNWLQIVMRSKCNHRTGFSQDP:SDC
exonB FGPTPSPTPSPTSTSQDP:TRSDAAHPLAELAAP:SF
```

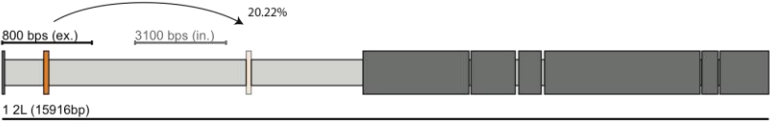
Gene: IA-2, IA-2 ortholog, FBgn0031294  
Polypeptide: IA-2-PC, FBpp0290630



For clarity introns have been scaled down by a factor of 6.81

```
.....10.....
exonA ATEIIFLLC-FYSHVCCFD
exonB GCQFVRTLCIP:HSFV-CYD
```

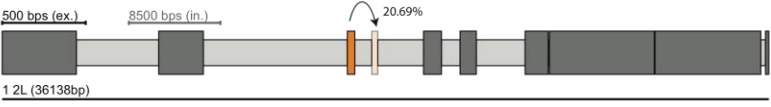
Gene: ush, u-shaped, FBgn0003963  
Polypeptide: ush-PA, FBpp0077723



For clarity introns have been scaled down by a factor of 3.82

```
.....10.....
exonA GDCSDTAEMTVDSR
exonB GWTETVEVHIELQ
```

Gene: CG32982, FBgn0052982  
Polypeptide: CG32982-PE, FBpp0290262

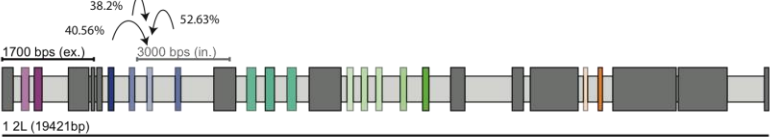


For clarity introns have been scaled down by a factor of 15.52

```
.....10.....
exonA VSCNKQTNNLNFQD
exonB IEC---TMWLDRRRES
```

Gene: Mhc, Myosin heavy chain, FBgn0264695  
Polypeptide: Mhc-PA, FBpp0080453

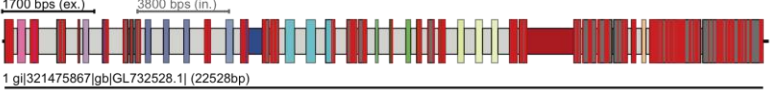
RNA-Seq data supports 3'-end of exon C.  
Verified by literature.  
Conserved in Aedes aegypti, Anopheles gambiae, Apis mellifera, Atta cephalotes, Daphnia pulex, Pediculus humanus corporis, Tribolium castaneum, dana, dere, dgri, dmoj, dper, dpse, dsec, dvir, dwil and dyak.



For clarity introns have been scaled down by a factor of 1.75

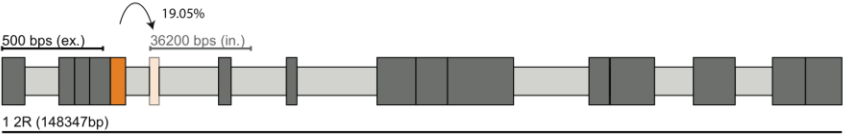
```
.....10.....20.....30.....
exonA DICLLTDNIYDYHIVSQKVTVASIDDAEEFSLTD
exonB EYCLLSNNIYDYRIVSQKTTIPSVNDGEEWVAVD
exonC EMVFLGQHIGDYPGICQKTRIPGVNDGEEFELTD
exonD EMCFLSDNIYDYNVSVQKVTIPNMDDGEEFQLAD
DapExonA ADCCLVDDIYQYNFVSQKITIPSMDDSEEMALTD
DapExonB ADCSLVDDIYTYNFVSQKITIPSMDDSEEMGLTN
DapExonC ADCRLVDDIYTYNFVSQKITIPSMDDNEEMGLTD
DapExonD AMCSLSDNIYDYFVSQKVTVP:SIDDSEEMQ:MD
```

Cross-species search in Daphnia pulex



For clarity introns have been scaled down by a factor of 2.25

Gene: Gprk1, G protein-coupled receptor kinase 1, FBgn0260798  
Polypeptide: Gprk1-PA, FBpp0110413



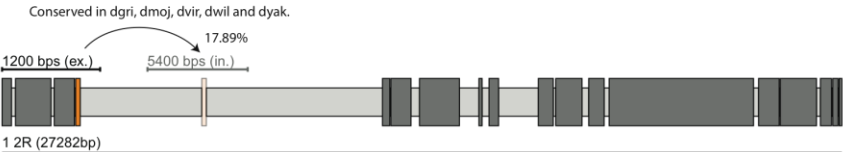
10 20  
exonA EYSKHAVA SVQKYL LKNEV PVDLFE  
exonB -----CKIFLLKNEVLVDLFE

Gene: CG30438, FBgn0050438  
Polypeptide: CG30438-PB, FBpp0085404



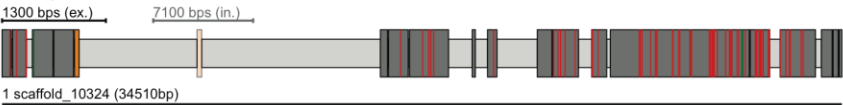
10 20  
exonA GGT KSHKIF FWE LAKGLISR  
exonB GGLPEETT RKNRVQ KQWSQ

Gene: brp, bruchpilot, FBgn0259246  
Polypeptide: brp-PD, FBpp0289193

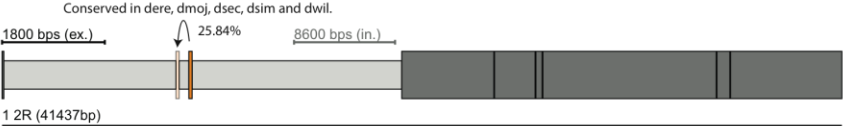


10 20  
exonA G--KEERQMFQQMQAMA-QKQ  
exonB ---EQEQNRTFDSIQKSI SQA  
dvirExonA G--KEERQMFQQMQAMA-QKQ  
dvirExonB GVKREKERRRRQMQCA--KQ

Cross-species search in dvir

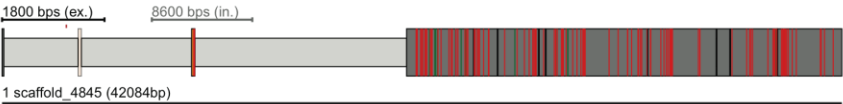


Gene: shn, schnurri, FBgn0003396  
Polypeptide: shn-PD, FBpp0089118



10 20  
exonA KTTIVIKC-SKWVTSRHQEK  
exonB KSTVN-SRKSALETAREKTK  
dereExonA KQQIKATHK-ANRNKTQKIK  
dereExonB KSTVN-SRKSALESVREKPK  
dmojExonA KSTVN-SRKNLTREKLEK  
dmojExonB KQQQLSKKKCLSSALESSK

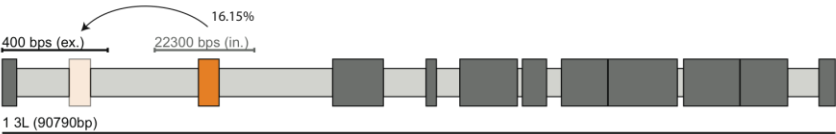
Cross-species search in dvir



For clarity introns have been scaled down by a factor of 4.87

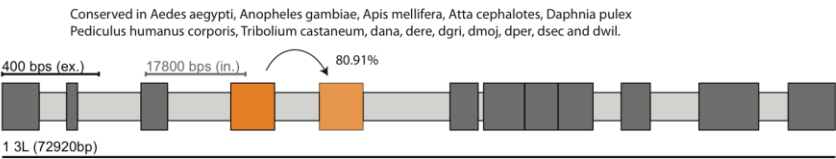


Gene: Eip63E, Ecdysone-induced protein 63E, FBgn0264001  
Polypeptide: Eip63E-PD, FBpp0072990

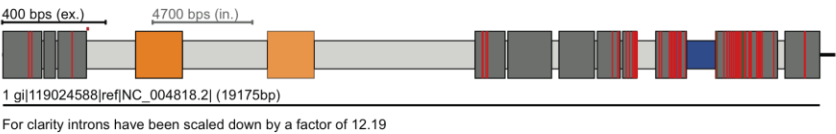


.....10.....20.....  
exonA GSTKIEKSDLKIQVIYMQMSNKYGQRG  
exonB GVTMRKKGGALQKLKKRLSHSFG-RL

Gene: nAcRalpha-80B, nicotinic Acetylcholine Receptor alpha 80B, FBgn0037212  
Polypeptide: nAcRalpha-80B-PC, FBpp0289395

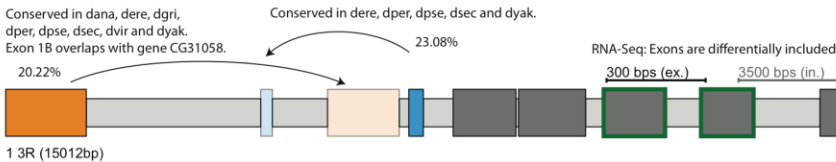


Cross-species search in *Anopheles gambiae*



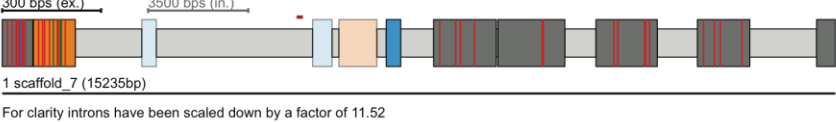
.....10.....20.....30.....40.....50.....60.....  
exonA ADGNFEVTLATKATIIYSEGLVENKPPAIYKSSCEIDVEYFPFDEQTCVLKFGSWTYDGFK  
exonB ADGHYEVTLMTKAIYVNNGLVIWQPPAVYKSSCSIDVEYFPYDVQTCILKLGSWTYDGFK  
AngExonA ADGNFEVTLATKATIIYSEGLVENKPPAIYKSSCEIDVEYFPFDEQTCVLKFGSWTYDGFK  
AngExonB ADGHYEVTLMTKATVYNNGMVIWQPPAVYKSSCSIDVEYFPYDVQTCVLKLGSWTYDGFK

Gene: tau, FBgn0051057  
Polypeptide: tau-PA, FBpp0084567

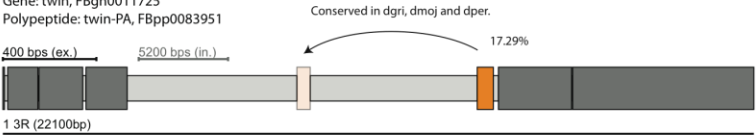


.....10.....20.....  
exonA VGDSDS---ESAQVA  
exonB EGDNDSGVDESTQEK  
dperExonA ELSNGFGPSQSQA  
dperExonB EQSDNGSAADEAGNAATAES  
dperExonC EGDNDSGVDESTQEK

Cross-species search in *dper*

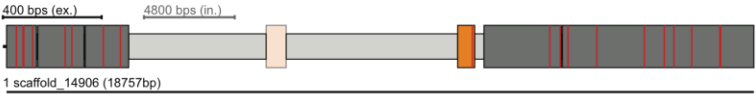


Gene: twin, FBgn0011725  
Polypeptide: twin-PA, FBpp0083951



For clarity introns have been scaled down by a factor of 13.60

Cross-species search in dgri

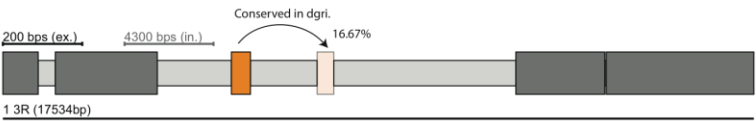


For clarity introns have been scaled down by a factor of 13.08

```

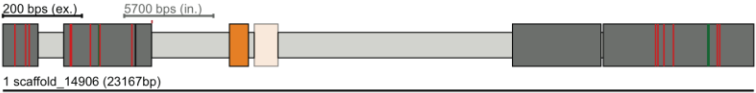
      10      20
exonA      FFQAP---PPL--WVPL--ENNPSEFW
exonB      FTVNP---PPQRFWLPFLAKPNKTRPA
dgriExonA  FAVTBSLTPPPLPSSPLSQAGHNRRP
dgriExonB  FTVNP---PPQRFWLPFLAKPNKSRPA
dmojExonA  FFYLPVRSTRPIA~QLQMRKPNKSRRLH
dmojExonB  FTVNP---PPQRFWLPFLAKPNKSRPA
```

Gene: abd-A, abdominal A, FBgn0000014  
Polypeptide: abd-A-PA, FBpp0082828



For clarity introns have been scaled down by a factor of 19.19

Cross-species search in dgri



For clarity introns have been scaled down by a factor of 25.25

```

      10      20
exonA      D---WMGSPFERVVCGDFN
exonB      D---W---RDFSSVVVGRQT
dgriExonA  D---WMGSPFERVVCGDFN
dgriExonB  DTSGNQPMFSSSLIVDFCN
```

Gene: CG14741, FBgn0037989  
Polypeptide: CG14741-PC, FBpp0297858



For clarity introns have been scaled down by a factor of 1.86

```

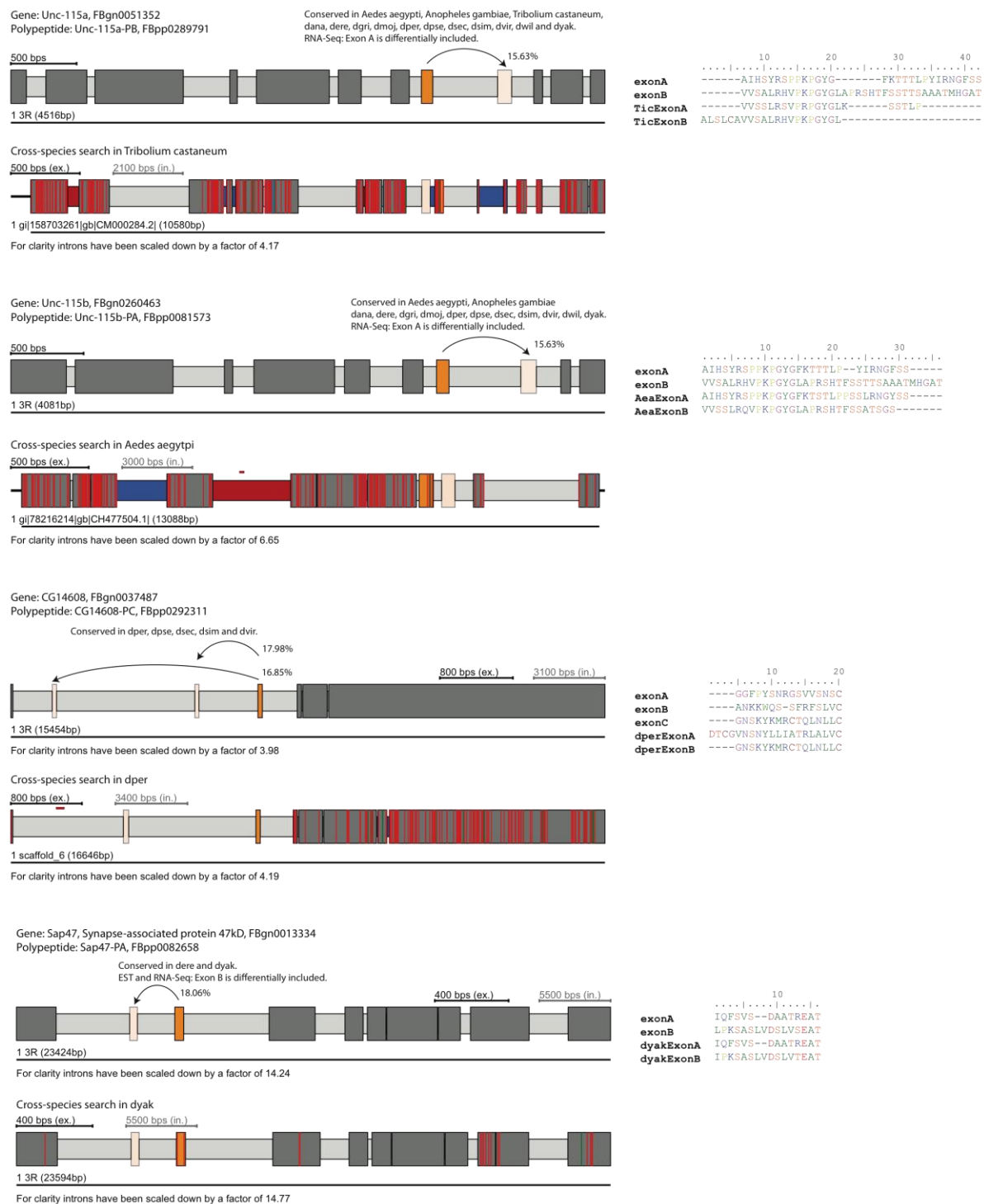
      10      20
exonA      EGEGPKRDHDDAFGTWHRKH
exonB      ENERRIRANDKEFNAQFKYH
```

Gene: CG6241, FBgn0037792  
Polypeptide: CG6241-PA, FBpp0081663

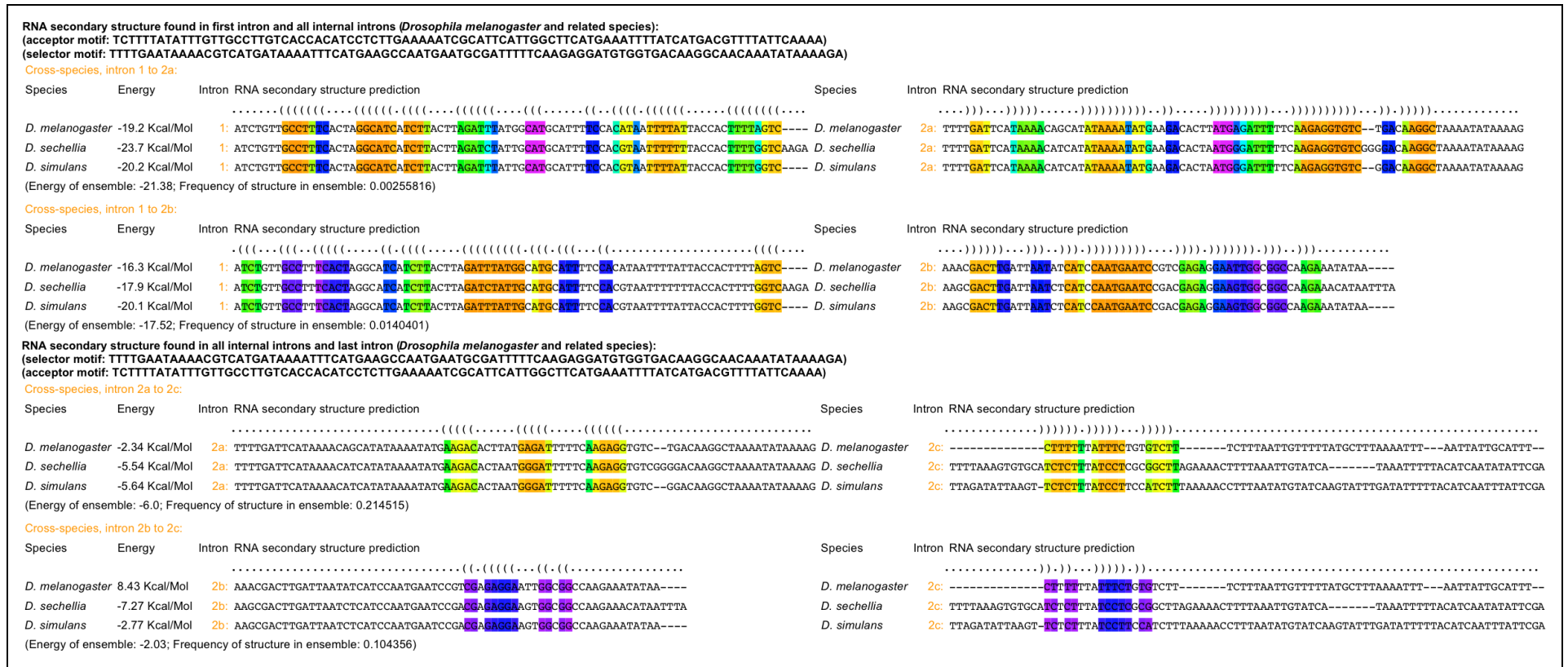


```

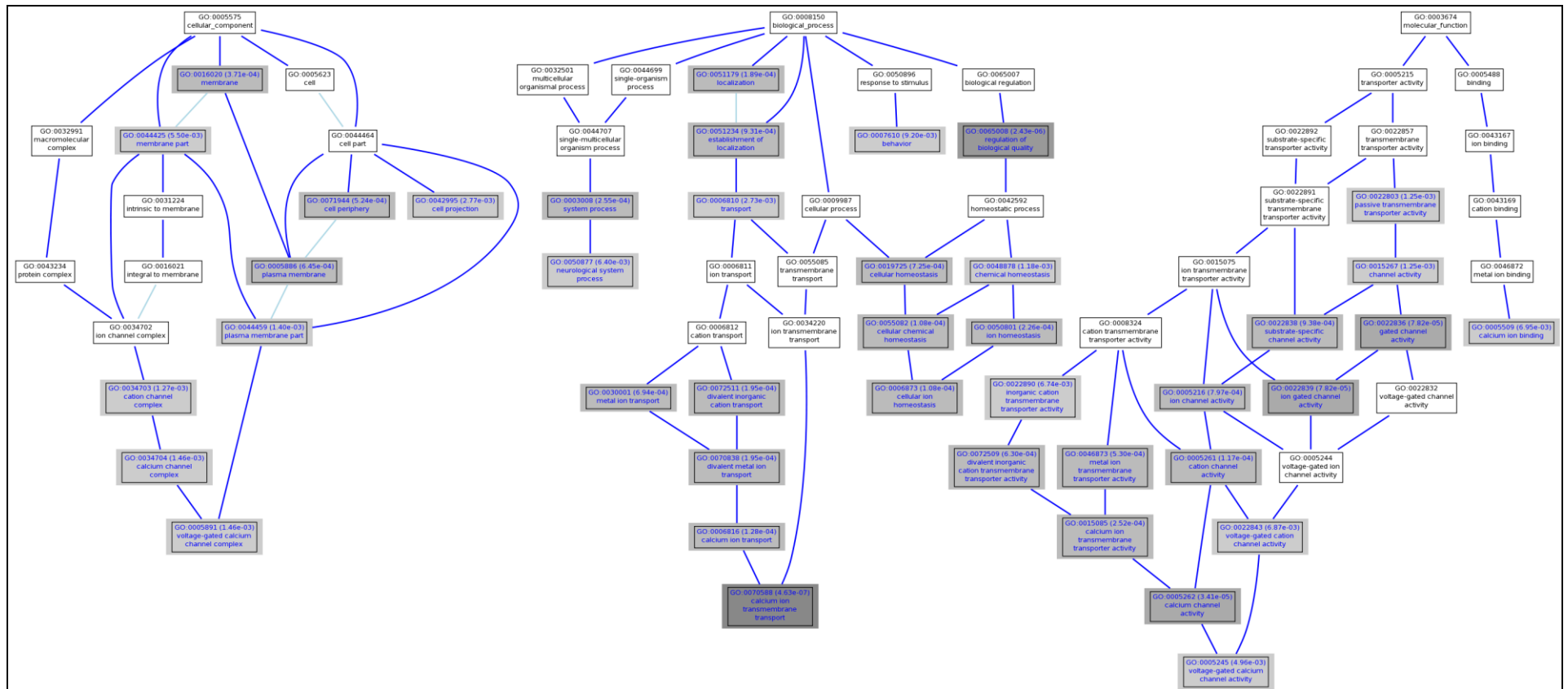
      10      20      30      40
exonA      DARIINYNHKTFKKGKKKSTLTGDPNDERAKFRLWNRTK
exonB      -----TLTGDPNDERAKFRLWNRTK
```



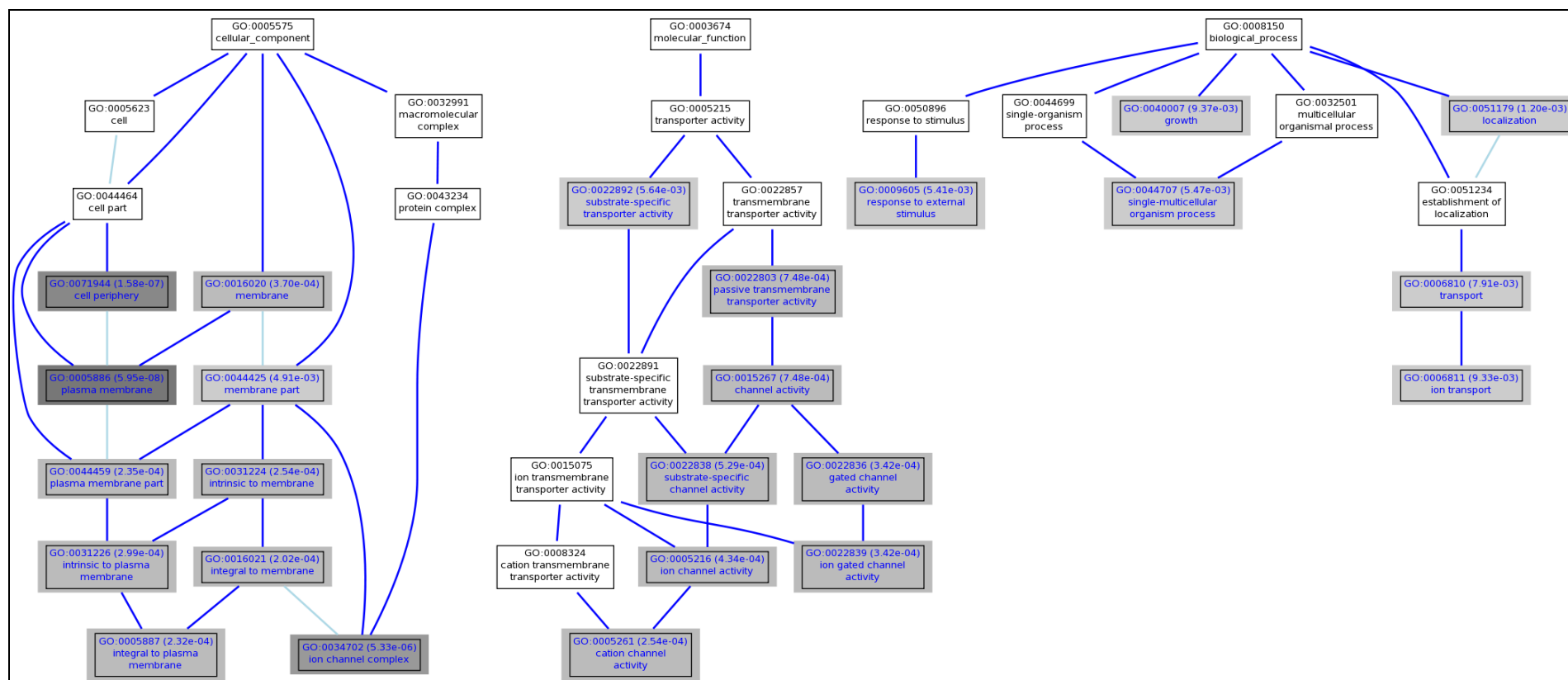
**Supplementary Figure S23. Genes containing newly predicted mutually exclusive spliced exons (MXEs) which were not annotated in Flybase release 5.36 nor in release 5.48.** All transcripts are represented 5' to 3'. The color coding is explained in the legend. Colored big bars represent MXEs. The darkest colored bar is the exon that was included in the query sequence, while the lighter colored bars represent identified MXEs. The higher the similarity between the candidate and the query exon the darker the color of the candidate (100% identity would result in the same color). The opacity of the colors of each alternative exon corresponds to the alignment score of the alternative exon to the original one.



**Supplementary Figure S24. RNA secondary structure prediction for gene CG14608 of *Drosophila melanogaster*.**



**Supplementary Figure S25. Gene Ontology (GO) term enrichment analysis of genes containing mutually exclusive spliced exons (MXEs), which are annotated and reconstructed.**



**Supplementary Figure S26. Gene Ontology (GO) term enrichment analysis of genes containing mutually exclusive spliced exons (MXEs), which were predicted but not annotated.**

## 2 Supplementary Tables

**Supplementary Table S1.** The table shows the numbers of mutually exclusive spliced exons (MXEs), which were annotated in Flybase release r5.36, which were predicted, and which were annotated and also predicted. For the prediction criteria a maximal length difference of 20 aa, a minimal similarity score of 15% and a minimal original exon length of 15 aa was used. Predicted MXE candidates, which overlap (and not exactly match) an already annotated exon, were filtered out.

		Matching prediction criteria of MXEs					
	Annotated MXEs	Annotated AND predicted MXEs		Predicted MXEs		Annotated as constitutive or differentially included	
			Cross / EST evidence		Cross / EST evidence		Annotated as MXEs in r5.48
Initial	660	31	28 / 17	65	47 / 20	2	0
3'-terminal	376	42	36 / 22	55	45 / 25	8	0
Internal	261	218	206 / 56	419	321 / 88	159	5
Sum	1297	291	270 / 95	539	413 / 133	169	5

**Supplementary Table S2.** The table shows the numbers of mutually exclusive spliced exons (MXEs), which were annotated in Flybase release r5.36, which were predicted, and which were annotated and also predicted. In contrast to Table S1, the minimal similarity score was set to 10% instead of 15% (the maximal length difference of 20 aa and the minimal original exon length of 15 aa are unchanged). Predicted MXE candidates, which overlap (and not exactly match) an already annotated exon, were filtered out.

		Matching prediction criteria of MXEs		
	Annotated MXEs	Annotated AND predicted MXEs	Predicted MXEs	Annotated as constitutive or differentially included
Initial	660	42	205	4
3'-terminal	376	48	106	10
Internal	261	228	844	198
Sum	1297	318	1155	212

**Supplementary Table S3.** The table shows the versions and sources of the genome sequence, protein annotation and EST datasets.

Species	Dataset release	URL
<i>Drosophila melanogaster</i>	dmel_r5.36_FB2011_04	ftp://ftp.flybase.net/genome
<i>Drosophila ananassae</i> TSC#14024-0371.13	dana_r1.3_FB2011_07	
<i>Drosophila erecta</i> TSC#14021-0224.01	dere_r1.3_FB2011_08	
<i>Drosophila grimshawi</i> TSC#15287-2541.00	dgri_r1.3_FB2010_02	
<i>Drosophila mojavensis</i> TSC#15081-1352.22	dmoj_r1.3_FB2011_05	
<i>Drosophila persimilis</i> MSH-3	dper_r1.3_FB2010_02	
<i>Drosophila pseudoobscura</i> MV2-25	dpse_r2.25_FB2011_10	
<i>Drosophila sechellia</i> Rob3c	dsec_r1.3_FB2011_08	
<i>Drosophila simulans</i> str. Mosaic	dsim_r1.3_FB2011_08	
<i>Drosophila virilis</i> TSC#15010-1051.87	dvir_r1.2_FB2011_07	
<i>Drosophila willistoni</i> TSC#14030-0811.24	dwil_r1.3_FB2010_02	
<i>Drosophila yakuba</i> Tai18E2	dyak_r1.3_FB2011_08	
<i>Daphnia pulex</i>	V1.0	http://www.ncbi.nlm.nih.gov/nuccore/ACJG000000000.1
<i>Anopheles gambiae</i> str. PEST	AgamP3	http://ftp.ncbi.nih.gov/genomes/Anopheles_gambiae
<i>Aedes aegypti</i> str. Liverpool	AaegL1	http://www.ncbi.nlm.nih.gov/nuccore/AAGE000000000.2
<i>Atta cephalotes</i>	Attacep1.0	http://www.ncbi.nlm.nih.gov/nuccore/ADTU000000000.1
<i>Apis mellifera</i> str. DH4	Amel_4.5	http://www.ncbi.nlm.nih.gov/nuccore/AADG000000000.6
<i>Tribolium castaneum</i> str. Georgia GA2	Tcas_3.0	http://www.ncbi.nlm.nih.gov/nuccore/AAJJ000000000.1
<i>Pediculus humanus corporis</i> str. USDA	JCVI_LOUSE_1.0	http://www.ncbi.nlm.nih.gov/nuccore/AAZO000000000.1
<i>Caenorhabditis elegans</i>	WS230	ftp://ftp.wormbase.org/pub/wormbase/releases
<i>Homo sapiens</i>	Build 37.3	ftp://ftp.ncbi.nih.gov/genomes/H_sapiens
<i>Arabidopsis thaliana</i>	TAIR10_genome_release	ftp://ftp.arabidopsis.org/home/tair/Genes
<i>Drosophila melanogaster</i> (EST data)	v2010_11_11	http://www.ncbi.nlm.nih.gov/nucest/?term=txid7227%5BOrganism%5D

### 3 Supplementary References

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50. Wilkin, M. B. *et al.* *Drosophila* dumpy is a gigantic extracellular protein required to maintain tension at epidermal-cuticle attachment sites. *Curr. Biol.* **10**, 559–567 (2000).