

Antagonistic action of Bicoid and the repressor Capicua determines the spatial limits of *Drosophila* head gene expression domains

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Bicoid (Bcd) is the anterior determinant in *Drosophila*. Accordingly, loss of Bcd causes loss of head and thorax and their replacement with posterior structures. *bcd* mRNA is maternally deposited at the anterior pole and Bcd forms an anterior-to-posterior (AP) concentration gradient. The expression of a series of zygotic head genes is thought to be differentially regulated by distinct threshold concentrations of the Bcd gradient. Thereby Bcd functions as a morphogen, instructing fields of cells to take on specific fates. Here, we show that spatial limits of anterior genes are also set in the absence of a Bcd gradient and depend on factors of the maternal terminal system. The receptor tyrosine kinase Torso (Tor), a key component of this system, is active in the pole regions of the embryo. Its activity downregulates the maternally deposited repressor Capicua (Cic), leaving high Cic activity in the central regions and decreasingly lower Cic activities toward the poles. We show that the positions of posterior boundaries of Bcd target genes are dependent not only on Bcd, but also on Tor-mediated Cic activity. The results indicate that Cic can mediate repression through distinct binding sites within a Bcd responsive enhancer and that gene activation by Bcd is antagonized by Cic. The activating and repressive effects of Bcd and Cic, respectively, are integrated by the Bcd target gene enhancer. We conclude that the spatial domains of head gene expression are determined by Bcd in concert with Tor-dependent repressors.

bicoid antagonist | *Drosophila* development | gene regulation | head development | morphogen gradient

Bcd is a homeodomain-containing transcription factor required for head development in *Drosophila* (1, 2). *bcd* mRNA is maternally deposited and localized to the anterior egg pole by its 3'-UTR (3, 4). From there both the mRNA and ultimately the protein form a concentration gradient along the anterior–posterior (AP) axis (5, 6). Interestingly, a change in *bcd* dosage leads to shifts in target gene expression. Reduction by one *bcd* copy leads to an anterior shift of target gene expression boundaries, whereas an additional copy results in a posterior shift (7). Thus, it has been suggested that Bcd functions as a morphogen (8, 9) and that target gene expression depends directly on distinct concentrations of Bcd along the AP axis (7). This concentration-dependent gene activation is thought to be mediated by the affinity of binding sites for Bcd within target genes (10–12). Targets expressed close to the source would contain low affinity binding sites, whereas targets far from the source would contain high affinity sites. Bcd also can activate enhancers via cooperative binding (13, 14).

In addition to Bcd, the terminal system has been shown to affect gene expression in the head region of the embryo. Torso (Tor), a receptor tyrosine kinase, is activated only at the poles (15) from where it signals through the MAP kinase pathway and regulates terminal gene expression by relief of repression (15–18). *capicua* (*cic*) mRNA is maternally deposited in the embryo, resulting in ubiquitous Cic expression. It has been shown that Tor downregulates the DNA binding repressor Cic at the termini by phosphorylation via the activated MAP kinase Rolled (19, 20), resulting in low Cic activity at the termini and high activity in the center of the

embryo. Loss of Cic leads to the derepression of head and tail genes and the expansion of these regions at the cost of the trunk and abdomen (19). Also, it has been suggested that Tor activity results in the phosphorylation of Bcd (21) strengthening its morphogenic nature along the AP axis (22, 23). At the anterior pole Tor has been proposed to have an opposite function, i.e., to downregulate Bcd activity, and that this effect causes repression of *hunchback* and *orthodenticle* at the anterior tip (21, 24).

Here we show that uniform expression of Bcd leads to ectopic head gene expression with mirror image polarity at the posterior pole. This effect is dependent on Tor-regulated Cic activity, confirming a major role of the terminal system in the spatial control of Bcd-dependent head gene expression. We found that Cic activity is necessary to determine the spatial limits of head gene expression by repression. These results suggest that this effect of the terminal system is mediated by binding sites located in Bcd responsive enhancers. We conclude that anterior patterning is dependent on the interpretation of activation by Bcd relative to repression by Cic by the enhancers.

Results

Uniform Expression of Bcd Causes the Mirror Image Duplications of Target Gene Expression. To assess the ability of Bcd to activate gene expression independent of its gradient, we used the UASp/Gal4 system (25, 26) to ectopically express *bcd* without its localizing 3'-UTR in the female germline. In embryos that derive from such females, the endogenous Bcd gradient was superimposed with transgene-derived Bcd, resulting in uniform Bcd levels in the posterior half of the embryo (*SI Materials and Methods* and *Fig. S1 A, B, D, and E*). In such embryos, the Bcd target gene *hunchback* (*hb*) (27), which is normally detected in the anterior 50% of the embryo, is ubiquitously expressed (*Fig. S2 A and B*). However, Bcd target genes that are normally confined to more anterior regions, such as *cap-n-collar* (*cnc*) (28), *tailless* (*tll*) (29), and *giant* (*gt*) (30) are expressed in distinct but ectopic domains (*Fig. 1*; see also *Figs. S2 C, D, G, and H* and *S3 A–C*). *cnc*, normally expressed only in the anterior region (28) (*Fig. 1 A and B*), was also expressed in the posterior of embryos, which contain uniform high levels of Bcd (*Fig. 1 E and F*). Similarly, the anterior tip domain of Gt expression (*Fig. 1 A and C*) was duplicated in the posterior (*Fig. 1 E and G*). Gt expression was also detected in a central, ventrolaterally repressed domain, which resembled the anterior portion of the anterior stripe in wild-type embryos. Finally, the dorsolateral anterior domain of *tll* (*Fig. 1 A and D*) was broadly expanded along

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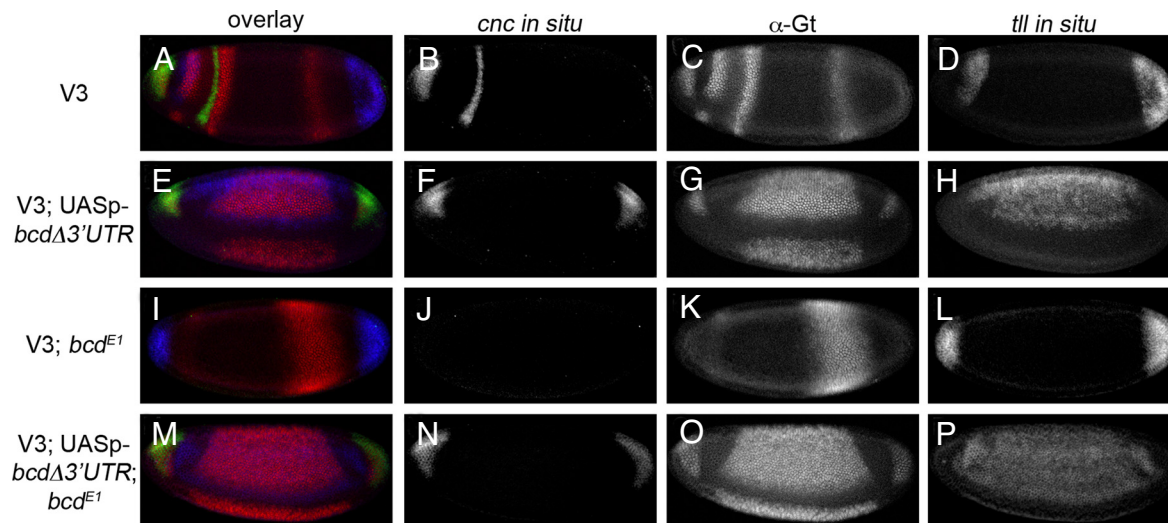


Fig. 1. Uniform expression of transgene-derived Bcd in wild-type and *bcd^{E1}* embryos causes mirror image duplications of anterior expression domains in the posterior region. *cnc* and *tll* mRNAs were detected by fluorescent *in situ* hybridization (green and blue, respectively); Gt was detected by immunohistochemistry (red); A, E, I, and M are overlays; B–D, F–H, J–L, and N–P are single channel gray scale images; anterior is to the Left; dorsal is Up. (A–D) Expression of *cnc*, Gt, and *tll* in a control embryo (V3). (A and B) *cnc* is expressed in an anterior cap and a more central collar. (A and C) Gt is expressed in an anterior tip domain, an anterior double stripe domain, consisting of a discontinuous and a continuous stripe, and a posterior stripe domain. (A and D) *tll* is expressed in an anterior dorsal-ventral and a posterior cap domain. (E–H) Transgene-derived ubiquitous Bcd causes mirror image duplications of anterior patterns in the posterior. (E and F) The *cnc* cap domain is duplicated at the posterior pole. (E and G) The tip domain of Gt is also duplicated at the posterior and Gt is expressed centrally as a ventrolaterally repressed stripe. This expression corresponds to the discontinuous anterior stripe. (E and H) *tll* is expressed in a broad, ventrally repressed domain in the center of the embryo. (I–L) Expression of *cnc*, Gt, and *tll* in *bcd^{E1}* embryos. (I and J) *cnc* expression is completely absent. (I and K) Gt is only detected as a broadened posterior stripe. (I and L) In the anterior *tll* is expressed in a cap, resembling posterior expression. (M–P) Uniform Bcd in *bcd^{E1}* embryos causes mirror image duplications. (M and N) The anterior cap of *cnc* expression is restored and duplicated in the posterior. (M and O) Gt tip expression is also restored and duplicated, while a ventrolaterally repressed stripe is expressed in the central regions. (M and P) *tll* is expressed in a central ventrally repressed domain. Note that the expression patterns observed in M–P strongly resemble those observed in E–H, showing that the mirror image duplications do not depend on the endogenous Bcd gradient.

the AP axis of the embryo (Fig. 1 E and H). These findings indicate that the anterior patterns of these three genes were mirrored along an axis, which runs vertically through the anterior *tll* domain, in an area in which it overlaps with the anterior Gt stripe. We confirmed that the ectopic expression domains observed for *cnc* and Gt were indeed mirror image duplications by the use of *lacZ* reporter constructs. The *cnc*(+5)-*lacZ* construct recapitulates *cnc* in the wild-type embryos (31) (Fig. S2E), while *gt*(-6)-*lacZ* recapitulates the *gt* tip expression (31) (Fig. S2I). In the presence of unlocalized Bcd both the *cnc*(+5) and the *gt*(-6) drive *lacZ* expression in distinct domains in the posterior (Fig. S2F and J) as was observed for *cnc* and Gt in the presence of unlocalized Bcd. Thus, uniformly expressed Bcd causes an expansion and mirroring of anterior expression domains at the posterior pole (Fig. 1E). In fact, the Hox gene *labial* (*lab*) (32), normally expressed in a stripe anterior to the cephalic furrow (33) (Fig. S2K), was duplicated in a mirror image fashion in the posterior of embryos (Fig. S2L), indicating that these cells have indeed taken on specific anterior identities. Additionally, we observed that the expression domains of other potential Bcd target genes in the head region, such as those of *knirps* (*kni*) (34), *orthodenticle* (*otd*) (12), *buttonhead* (*btd*) (35), *empty spiracles* (*ems*) (36), and *sloppy-paired* (*slp2*) (37), are duplicated with mirror image polarity in the posterior region in the presence of uniform Bcd (Fig. S3).

To exclude that the endogenous Bcd gradient caused the observed mirror image duplications, we examined the effects of uniform Bcd in embryos lacking endogenous Bcd. In the absence of endogenous Bcd no gradient is detectable in the presence of ectopic unlocalized Bcd, indicating that all nuclei in the embryo receive a similar amount of Bcd (Fig. S1C and F). In embryos from females homozygous for the *bcd^{E1}* loss-of-function allele (2), the anterior domains of *cnc* and Gt are lost and anterior *tll* expression strongly resembles its posterior expression (28–30, 38) (Fig. 1 I–L). In such embryos, uniform Bcd caused mirror image expression of

head genes indistinguishable from its effect in the presence of endogenous Bcd (compare E–H and M–P in Fig. 1). *cnc* expression was restored at both poles of the embryos (Fig. 1 M and N). Thus, endogenous Bcd gradient information cannot be responsible for residual anterior gene expression and their mirror image expression patterns in the posterior. This result shows that although Bcd is necessary to activate head genes such as *cnc*, differential activation of target genes and their spatial order is not dependent on Bcd gradient information. Similar effects have been observed in embryos that express low levels of Bcd uniformly along the entire axis (11, 22, 39).

Terminal System Activity Is Required for the Mirror Image Duplications of Head Gene Expression Domains. It has been shown that the maternal terminal system activates anterior target genes by relief of repression (16, 17, 19). One of the main effectors of Tor signaling is Cic, a ubiquitous repressor of anterior and posterior gene expression that is downregulated at the embryonic termini by activated Tor signaling (19). Consequently, Cic and/or other Tor-dependent repressors could antagonize Bcd-dependent gene activation and thereby position posterior boundaries (PBs) of Bcd target genes in the head. As Tor signals at both termini, Cic could also repress Bcd-dependent target genes in embryos which have received uniform levels of Bcd, causing the observed mirror image duplications of anterior expression domains in the posterior region.

In embryos from females homozygous for the *cic¹* loss-of-function allele, the head and tail regions are expanded at the expense of the trunk (19). Consequently, *cnc*, Gt, and *tll* expression is expanded toward the center (19) (Fig. 2 A–D), indicating that although their anterior domains were delimited, their PBs were not properly positioned (see below). In *cic¹* embryos uniformly expressing Bcd, *cnc* expression domains appeared at both termini and were connected via a ventrolateral stripe (Fig. 2 E and F). Gt was also expressed in both pole regions (Fig. 2 E and G) and *tll* expression

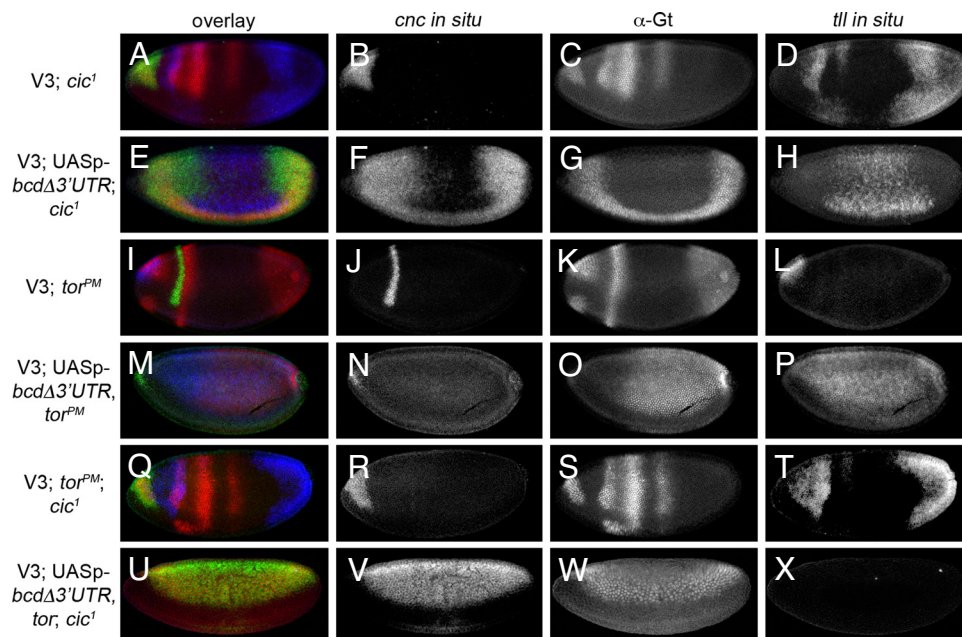


Fig. 2. The terminal system components Cic and Tor are required for mirror image duplications in response to uniformly expressed Bcd. Gene expression was visualized as described in Fig. 1. *A, E, I, M, Q, and U* are overlays of all three channels; *B–D, F–H, J–L, N–P, R–T, and V–X* are single channel gray scale images; anterior is to the *Left*; dorsal is *Up*. (*A–D*) Expression of *cnc*, *Gt*, and *tll* in *cic*¹ embryos. (*A and B*) The cap domain of *cnc* is expanded toward the center and the collar is missing; compare to Fig. 1*B*. (*A and C*) Main domains of *Gt* expression are formed, but both stripe domains are ventrally repressed. The posterior *Gt* stripe is shifted toward the center. (*A and D*) The anterior and posterior expression domains of *tll* are expanded toward the center. (*E–H*) Ubiquitous Bcd in *cic*¹ embryos causes duplications with minimal patterning information. (*E and F*) The cap domain of *cnc* is expanded and duplicated at the posterior, and both caps are connected by a ventrolateral stripe of *cnc* expression. (*E and G*) Anterior tip expression of *Gt* is expanded and duplicated. Both domains are connected by a ventrolateral stripe. (*E and H*) *tll* is expressed in the central region but not on the ventral side. This amounts to a duplication of only the anteriormost region with very limited positional information. (*I–L*) Expression of *cnc*, *Gt*, and *tll* in *tor*^{PM} embryos. (*I and J*) The *cnc* cap domain is absent; the collar is shifted toward the anterior. (*I and K*) The anterior tip domain of *Gt* is absent, the anterior *Gt* stripe domain is shifted to the anterior pole region and the expanded posterior *Gt* stripe is shifted to the posterior pole. (*I and L*) *tll* expression is absent in the posterior and the anterior dorsal-ventral wedge of expression is shifted to the anterior pole. (*M–P*) Uniformly expressed Bcd in *tor*^{PM} embryos does not cause duplications, but is unable to activate *cnc*. (*M and N*) *cnc* expression is only detected in a few anterior cells, indicating that it is strongly repressed even in the presence of excess Bcd. (*M and O*) Anterior tip expression of *Gt* is not recovered, but a broad ventrolaterally repressed, continuous *Gt* domain is observed, which most likely corresponds to the anterior discontinuous stripe. (*M and P*) *tll* is expressed throughout the embryo and overlaps with *Gt* expression. Note that *tll* expression is not excluded from the posterior pole. (*Q–T*) Expression of *cnc*, *Gt*, and *tll* in *tor; cic*¹ embryos resembles expression in *cic*¹ embryos (*A–D*). (*Q and R*) *cnc* expression at the anterior is restored in these embryos (compare to *I and J*) and the collar is missing. (*Q and S*) *Gt* expression strongly resembles *Gt* expression in *cic*¹ embryos (*A and C*) as does *tll* expression (compare *Q and T* to *A and D*). (*U–X*) Ubiquitous Bcd in *tor; cic*¹ embryos does not cause duplications, but continuous expression of the anteriormost targets. (*U and V*) *cnc* is expressed from the anterior to the posterior tip in a continuous, ventrally repressed domain. (*U and W*) *Gt* is detected in a ventrolateral stripe spanning the entire embryo, most likely corresponding to an extremely elongated tip domain. (*U and X*) *tll* is absent in these embryos. Thus, all cells resemble the anterior tip of a wild-type embryo.

was confined to dorsolateral regions, excluding *cnc*- or *Gt*-expressing cells (Fig. 2 *E and H*). These patterns were distinctly different from those observed upon ubiquitous Bcd expression in wild-type or *bcd*^{E1} embryos (compare Fig. 2 *E–H* with Fig. 1 *E–H* and *M–P*), indicating that the removal of Cic activity resulted in the expansion of Bcd target gene expression toward the center. However, as minimal anterior patterning was observed at both poles in *cic*¹ embryos expressing uniform Bcd (i.e., *cnc* and *Gt* expression at both poles separated by *tll* expression), positional information must be provided by additional factors.

To test whether such additional factors are also under the control of the terminal system, we examined the effects of ubiquitous Bcd in embryos lacking Tor activity. Such embryos, derived from *tor*^{PM} homozygous females, fail to develop head and tail structures (15). Because of ectopic Cic activity in the terminal regions of *tor*^{PM} embryos (19), the more central expression domains are shifted toward the termini, i.e., *cnc* and *Gt* expression are lost from the anterior tip and both anterior *Gt* stripe and *tll* are shifted to the anterior pole (28–30) (Fig. 2 *I–L*). Uniform Bcd expression in *tor*^{PM} embryos caused no duplications (Fig. 2 *M–P*), but embryos were continuously patterned, i.e., *Gt* and *tll* were expressed in overlapping, ventrally repressed domains throughout the embryo. However, *cnc*, a marker of anteriormost gene expression, was only

weakly restored in some embryos. Thus, in embryos lacking Tor activity, genes normally expressed at the anterior tip of the embryo were repressed despite the presence of Bcd. Thus the entire embryo was continuously patterned, but the anteriormost information (i.e., *cnc* expression) was missing. Corresponding results were reported from embryos containing low uniform levels of Bcd in the absence of *tor* (22).

We next expressed Bcd in embryos devoid of both *tor* and *cic* to observe possible Cic-independent effects of Tor. Overall, expression of target genes in embryos devoid of both *tor* and *cic* (Fig. 2 *Q–T*) was very similar to that in *cic*¹ embryos (19) (Fig. 2 *A–D*). Upon uniform Bcd expression, *cnc* was expressed in a dorsolateral domain along the entire length of the embryo (Fig. 2 *U and V*), *Gt* was expressed in a horizontal stripe of cells (Fig. 2 *U and W*), and *tll* expression was undetected (Fig. 2 *U and X*). Thus, the anterior pattern was not duplicated, but instead the entire embryo resembled the anterior tip region. In summary, uniformly expressed Bcd can cause anterior gene expression throughout the entire embryo only in the absence of two key components of the terminal system. These results indicate that (*i*) terminal Tor signaling is necessary to establish mirror image duplications in the presence of Bcd in the posterior pole region and (*ii*) Tor does not act in this process through Cic alone. In the presence of uniform Bcd, positional

much higher levels in the anterior of the embryo than necessary for proper gene activation, we have found that Bcd is not able to precisely position the PBs of head genes in the absence of Cic. In summary, we conclude that Bcd is not a “classical morphogen” as initially defined by Wolpert (8), but rather represents the activating component of a maternal “morphogenic network” that includes the terminal system. This network is required to set up both anterior–posterior polarity and to determine the spatial limits of gene expression in the head region of *Drosophila* embryos.

Materials and Methods

The following mutant alleles were used: *w¹¹¹⁸*, for *P*-element transformation and as wild-type reference strain; *bcd^{E1}* (*bcd⁶*), *tor^{WK}* (*tor¹*), *tor^{PM}* (*tor⁴*), and *cic¹*. *bcd* cDNA was cloned into UASp without its 3′-UTR (UASp-*bcd*Δ3′ UTR)

and expressed with V3-Gal4 (in figures and figure legends referred to as V3) (51) in the female germline. The *bcd*3T enhancer was cloned with or without *tor*REs into pCaSpeR-hs43-*lacZ* containing *attB* sites. *attB* vectors were injected into embryos from females carrying the ϕ C31-integrase on chromosome IV and an *attP* landing site on chromosome III at 86Fb (line ZH-*attP*-86Fb) (52). Immunohistochemistry and *in situ* hybridizations were conducted using standard methods. See *SI Materials and Methods* for additional information.

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- Berleth T, et al. (1988) The role of localization of *bicoid* RNA in organizing the anterior pattern of the *Drosophila* embryo. *EMBO J* 7:1749–1756.
- Frohnhofer HG, Nüsslein-Volhard C (1986) Organization of anterior pattern in the *Drosophila* embryo by the maternal gene *bicoid*. *Nature* 324:120–125.
- St Johnston D, Nüsslein-Volhard C (1992) The origin of pattern and polarity in the *Drosophila* embryo. *Cell* 68:201–219.
- Macdonald PM, Struhl G (1988) *cis*-acting sequences responsible for anterior localization of *bicoid* mRNA in *Drosophila* embryos. *Nature* 336:595–598.
- Spirov A, et al. (2009) Formation of the *bicoid* morphogen gradient: An mRNA gradient dictates the protein gradient. *Development* 136:605–614.
- Driever W, Nüsslein-Volhard C (1988) A gradient of *bicoid* protein in *Drosophila* embryos. *Cell* 54:83–93.
- Driever W, Nüsslein-Volhard C (1988) The *bicoid* protein determines position in the *Drosophila* embryo in a concentration-dependent manner. *Cell* 54:95–104.
- Wolpert L (1969) Positional information and the spatial pattern of cellular differentiation. *J Theor Biol* 25:1–47.
- Gurdon JB, Bourillot PY (2001) Morphogen gradient interpretation. *Nature* 413:797–803.
- Driever W, Thoma G, Nüsslein-Volhard C (1989) Determination of spatial domains of zygotic gene expression in the *Drosophila* embryo by the affinity of binding sites for the *bicoid* morphogen. *Nature* 340:363–367.
- Struhl G, Struhl K, Macdonald PM (1989) The gradient morphogen *bicoid* is a concentration-dependent transcriptional activator. *Cell* 57:1259–1273.
- Gao Q, Finkelstein R (1998) Targeting gene expression to the head: The *Drosophila* orthodenticle gene is a direct target of the Bicoid morphogen. *Development* 125:4185–4193.
- Rivera-Pomar R, Lu X, Perrimon N, Taubert H, Jäckle H (1995) Activation of posterior gap expression in the *Drosophila* blastoderm. *Nature* 376:253–256.
- Lebrecht D, et al. (2005) Bicoid cooperative DNA binding is critical for embryonic patterning in *Drosophila*. *Proc Natl Acad Sci USA* 102:13176–13181.
- Li WX (2005) Functions and mechanisms of receptor tyrosine kinase Torso signaling: Lessons from *Drosophila* embryonic terminal development. *Dev Dyn* 232:656–672.
- Liaw GJ, et al. (1995) The Torso response element binds GAGA and NTF-1/Elf-1, and regulates *tailless* by relief of repression. *Genes Dev* 9:3163–3176.
- Paroush Z, Wainwright SM, Ish-Horowitz D (1997) Torso signalling regulates terminal patterning in *Drosophila* by antagonising Groucho-mediated repression. *Development* 124:3827–3834.
- Furriols M, Casanova J (2003) In and out of Torso RTK signalling. *EMBO J* 22:1947–1952.
- Jiménez G, Guichet A, Ephrussi A, Casanova J (2000) Relief of gene repression by Torso RTK signaling: Role of *capicua* in *Drosophila* terminal and dorsoventral patterning. *Genes Dev* 14:224–231.
- Astigarraga S, et al. (2007) A MAPK docking site is critical for downregulation of *Capicua* by Torso and EGFR RTK signaling. *EMBO J* 26:668–677.
- Ronchi E, Treisman J, Dostatni N, Struhl G, Desplan C (1993) Down-regulation of the *Drosophila* morphogen Bicoid by the Torso receptor-mediated signal transduction cascade. *Cell* 74:347–355.
- Gao Q, Wang Y, Finkelstein R (1996) *Orthodenticle* regulation during embryonic head development in *Drosophila*. *Mech Dev* 56:3–15.
- Janody F, Sturny R, Catala F, Desplan C, Dostatni N (2000) Phosphorylation of Bicoid on MAP-kinase sites: Contribution to its interaction with the torso pathway. *Development* 127:279–289.
- Bellaïche Y, Bandyopadhyay R, Desplan C, Dostatni N (1996) Neither the homeodomain nor the activation domain of Bicoid is specifically required for its down-regulation by the Torso receptor tyrosine kinase cascade. *Development* 122:3499–3508.
- Brand AH, Perrimon N (1993) Targeted gene expression as a means of altering cell fates and generating dominant phenotypes. *Development* 118:401–415.
- Rørth P (1998) Gal4 in the *Drosophila* female germline. *Mech Dev* 78:113–118.
- Tautz D (1988) Regulation of the *Drosophila* segmentation gene hunchback by two maternal morphogenetic centres. *Nature* 332:281–284.
- Mohler J (1993) Genetic regulation of CNC expression in the pharyngeal primordia of *Drosophila* blastoderm embryos. *Roux's Arch Dev Biol* 202:214–223.
- Pignoni F, Steingrimsson E, Lengyel JA (1992) *bicoid* and the terminal system activate *tailless* expression in the early *Drosophila* embryo. *Development* 115:239–251.
- Eldon ED, Pirrotta V (1991) Interactions of the *Drosophila* gap gene *giant* with maternal and zygotic pattern-forming genes. *Development* 111:367–378.
- Schroeder MD, et al. (2004) Transcriptional control in the segmentation gene network of *Drosophila*. *PLoS Biol* 2:E271.
- Mlodzik M, Fjose A, Gehring WJ (1988) Molecular structure and spatial expression of a homeobox gene from the *labial* region of the *Antennapedia*-complex. *EMBO J* 7:2569–2578.
- Diederich RJ, Merrill VK, Pultz MA, Kaufman TC (1989) Isolation, structure, and expression of *labial*, a homeotic gene of the *Antennapedia* complex involved in *Drosophila* head development. *Genes Dev* 3:399–414.
- Rothe M, Wimmer EA, Pankratz MJ, González-Gaitán M, Jäckle H (1994) Identical transacting factor requirement for *knirps* and *knirps-related* gene expression in the anterior but not in the posterior region of the *Drosophila* embryo. *Mech Dev* 46:169–181.
- Wimmer EA, Simpson-Brose M, Cohen SM, Desplan C, Jäckle H (1995) Trans- and cis-acting requirements for blastodermal expression of the head gap gene *buttonhead*. *Mech Dev* 53:235–245.
- Hartmann B, Reichert H, Walldorf U (2001) Interaction of gap genes in the *Drosophila* head: *tailless* regulates expression of *empty spiracles* in early embryonic patterning and brain development. *Mech Dev* 109:161–172.
- Grossniklaus U, Cadigan KM, Gehring WJ (1994) Three maternal coordinate systems cooperate in the patterning of the *Drosophila* head. *Development* 120:3155–3171.
- Rivera-Pomar R, Jäckle H (1996) From gradients to stripes in *Drosophila* embryogenesis: Filling in the gaps. *Trends Genet* 12:478–483.
- Ochoa-Espinosa A, Yu D, Tsigiris A, Struffi P, Small S (2009) Anterior-posterior positional information in the absence of a strong Bicoid gradient. *Proc Natl Acad Sci USA* 106:3823–3828.
- Kawamura-Saito M, et al. (2006) Fusion between CIC and DUX4 up-regulates PEA3 family genes in Ewing-like sarcomas with t(4;19)(q35;q13) translocation. *Hum Mol Genet* 15:2125–2137.
- Li XY, et al. (2008) Transcription factors bind thousands of active and inactive regions in the *Drosophila* blastoderm. *PLoS Biol* 6:E27.
- Walldorf U, Kiewe A, Wickert M, Ronshaugen M, McGinnis W (2000) *Homeobrain*, a novel paired-like homeobox gene is expressed in the *Drosophila* brain. *Mech Dev* 96:141–144.
- Hahn M, Jäckle H (1996) *Drosophila* goosecoid participates in neural development but not in body axis formation. *EMBO J* 15:3077–3084.
- Russell SR, Sanchez-Soriano N, Wright CR, Ashburner M (1996) The *Dichaete* gene of *Drosophila melanogaster* encodes a SOX-domain protein required for embryonic segmentation. *Development* 122:3669–3676.
- Chen YJ, Chiang CS, Weng LC, Lengyel JA, Liaw GJ (2002) *Tramtrack69* is required for the early repression of *tailless* expression. *Mech Dev* 116:75–83.
- Florence BL, Faller DV (2008) *Drosophila* Female Sterile (1) Homeotic is a multifunctional transcriptional regulator that is modulated by Ras signaling. *Dev Dyn* 237:554–564.
- Schaeffer V, Killian D, Desplan C, Wimmer EA (2000) High *bicoid* levels render the terminal system dispensable for *Drosophila* head development. *Development* 127:3993–3999.
- Ochoa-Espinosa A, et al. (2005) The role of binding site cluster strength in Bicoid-dependent patterning in *Drosophila*. *Proc Natl Acad Sci USA* 102:4960–4965.
- Segal E, Raveh-Sadka T, Schroeder M, Unnerstall U, Gaul U (2008) Predicting expression patterns from regulatory sequence in *Drosophila* segmentation. *Nature* 451:535–540.
- Coppey M, Boettiger AN, Berezhkovskii AM, Shvartsman SY (2008) Nuclear trapping shapes the terminal gradient in the *Drosophila* embryo. *Curr Biol* 18:915–919.
- Häcker U, Perrimon N (1998) *DRhoGEF2* encodes a member of the Dbl family of oncogenes and controls cell shape changes during gastrulation in *Drosophila*. *Genes Dev* 12:274–284.
- Bischof J, Maeda RK, Hediger M, Karch F, Basler K (2007) An optimized transgenesis system for *Drosophila* using germ-line-specific ϕ C31 integrases. *Proc Natl Acad Sci USA* 104:3312–3317.